Res. 17(1): 81-83,2004

ANTIFUNGAL ACTIVITIES OF TRADITIONAL MEDICINAL PLANT EXTRACTS - A PRELIMINARY SURVEY

G. PERUMAL, CHEVULA SUBRAMANYAM, D. NATARAJAN, K. SRINIVASAN, C. MOHANASUNDARI and K. PRABAKAR*

Department of Microbiology, Kandaswami Kandar's College, Velur - 638 182, Tamil Nadu, India. *Department of Zoology, Jamal Mohamed College, Tiruchirappalli - 620 020, Tamil Nadu, India.

The purpose of this investigation was to establish the antifungal activities of ethanolic extract of nine medicinal plants against *Aspergillus flavus*, *A. terreus* and *Mucor* species. All the extracts in different concentrations exhibited good fungicidal properties.

Keywords: Antifungal activity; Ethanol extracts; Medicinal plants.

Inditional medicine is an important source of potentially useful new compounds for the development of chemotherapeutic agents1. The first step towards this goal is the screening of plants used in popular medicine. Thus, antimicrobial research is geared towards the discovery and development of novel antibacterial and antifungal agents. Because of the side effects and the resistance that pathogenic micro organisms build against antibiotics, much recent attention has been paid to extracts and biologically active compounds isolated from plant species used in herbal medicine². Plant based antimicrobials represent a vast untapped source for medicines and further exploration of plant, antimicrobials need to occur. Antimicrobials of plant origin have enormous therapeutic potential. They are effective in the treatment of infectious diseases while simultaneously mitigating many of the side effects that are often associated with synthetic antimicrobials3. Hence, the present study was carried out to investigate antifungal activities of ethanolic extracts of nine medicinal plants.

Source of fungi : The fungi used in this study are Aspergillus flavus, A. terreus and Mucor sp. They were obtained from Microbial Type Culture Collection (MTCC), IMTECH, Chandigarh and cultures were maintained on Sabouraud Dextrose Agar (SDA) plates for the further use.

Extract preparation : The method was followed as per Kumar and Prasad⁴ with slight alterations. Fresh, healthy plant parts such as Coleus aromaticus (leaves), Euphorbia tirucalli (stem), Trigonella foenum-graecum (leaves), Curcuma longa (rhizome), Aloe vera (aerial parts), Allium sativum (stem), Zingiber officinale (rhizome), Centella asiatica (leaves) and Solanum trilobatum (leaves) were collected and processed for the study. Ten grams of each plant material was weighed and surface sterilized with 0.1% Mercuric chloride and washed 3 times with distilled water. Then the material was grinded in pestle and mortar with 50% Ethanol. The extracts were filtered using Whattman No.1 filter paper and then centrifuged at 5000rpm for 5 minutes and finally evaporated. The residue was collected and it was made up to 40ml using distilled water. Further the extract was diluted to 25%, 50%, 75% and 100% concentrations.

Antifungal assay: About 15ml of SDA medium with 5ml of extract of different concentrations was poured into each sterilized petridishes. A set of three plates (replicates) was used in each concentration of extract and test organisms. The plates were gently shaked for the thorough mixing of the medium and the extract. After solidification of the medium, the test fungi were aseptically inoculated at the centre of the plate. From the 4th day onwards the fungal growth was observed in various concentrations of extracts and control. At the end of 10th day, results were recorded and tabulated.

Ethanolic extracts of nine traditional medicinal plants were screened for their antifungal activity against the fungal pathogens namely *Aspergillus flavus*, *A. terreus* and *Mucor* species were given in the table 1.

Among these extracts E. tirucalli and C. longa exhibited significant antifungal activity against A. flavus whereas other plant extracts found to be less active. But in case of T. foenum-graecum, C. longa, A. vera, C. asiatica and S. trilobatum, they produced maximum growth inhibition of A. terreus. All the other extracts showed minimum inhibitory activity against the test fungus. Similarly Satya et al.5 has proved that alcoholic extracts of Memecylon umbellatum showed slight antifungal activity including A. flavus. Abubacker and Ramanathan⁶ studied that the effect of Euphorbia splendens and Leonotis nepetaefolia extracts showed the maximum inhibitory activity against the Aspergillus species on varying concentrations of the extracts. Natarajan and Francis Xavier⁷ reported that the antifungal activity of ethanol and chloroform extracts from Tinospora cordifolia, showed good growth inhibition of A. niger and A, flavus. Like wise Alade and Irobl⁸ who showed that

S.No	Name of the plant	Concentration of extract (%)	Growth inhibition A. flavus	A. terreus	Mucor sp
1.	Coleus aromaticus	Control 25 50 75 100	++++ +++ ++ + +	++++ +++ ++ +	++++ ++ + - -
2.	Euphorbia tirucalli	Control 25 50 75 100	++++ ++ ++ -	++++ +++ ++ +	++++ +++ + -
3.	Trigonella foenum-graecum	Control 25 50 75 100	++++ +++ ++ + + + +	++++ ++ - - -	++++ +++ ++ -
4.	Curcuma longa	Control 25 50 75 100	++++ +++ + -	++++ + - -	+++++ + + -
5.	Aloe vera	Control 25 50 75 100	++++ +++ +++ ++- -	++++ ++ - -	++++ ++++ +++ +++ +++
6.	Allium sativum	Control 25 50 75 100	++++ ++++ +++ ++ +++ ++	++++ ++ + -	++++ ++ - -
7.	Zingiber officinale	Control 25 50 75 100	++++ +++ ++ +	++++ ++ - -	++++ ++ - - -
8.	Centella asiatica	Control 25 50 75 100	++++ ++++ +++ ++ +	++++ ++ - -	++++
9.	Solanum trilobatum	Control 25 50 75 100	++++ +++ +++ -	++++ ++ - -	++++ ++ -
+++	- maximum growth	++ - 50% ar	owth inhihiting	1000/	

Table1. Antifungal activities of ethanolic extracts of traditional medicinal plants.

+++ - 25% growth inhibition

++ - 50% growth inhibition + - 75% growth inhibition

100% growth inhibition
(or) no growth

the alcoholic extracts of *Acalypha wilkesiana* has antifungal activity against *A. flavus*.

In contrast, *C. longa* was found to be more effective against *Mucor* sp. by inhibiting the growth even at 25% concentration onwards, which is followed by *A. sativum, Z. officinale, C. asiatica* and *S. trilobatum* extracts. The extract from *A. vera* showed poor response against the test fungus. Irobi and Adedayo⁹ reported that extracts of *Hyphaene thebaicia*, was found to produce significant antifungal activity of *Mucor* sp. at a concentration between 3.1 to 25% of the extract.

The present and preliminary investigation was tested out *in vitro* screening of ethanolic extracts of nine medicinal plants against the growth of three pathogenic fungi. However, this study recommends that further phytochemical analysis of these medicinal plants for the role of active compounds used to cure diseases.

Acknowledgements

Authors wish to thank Management and Principal, Kandaswami Kandar's College, Velur for their support and the laboratory facilities used during the course of this investigation.

References

- Racio MC Rios JC and Villar A 1989, A review of some antimicrobial compounds isolated from medicinal plants reported in the literature 1978 - 1988. *Phytotherapy Research* 3(4) 117 - 125.
- 2. Essawi T and Srour M 2000, Screening of some Palestinian medicinal plants for antibacterial activity.

Journal of Ethnopharmocology 10 343-349.

- 3. Iwu M W, Duncan AR and Okunji C O 1999, New antimicrobials of plant origin. *In* : Janick.J.(Ed.), *Perspectives on New Crops and New Uses*. ASHS press. Alexandria, UA. PP.457-462.
- 4. Kumar S and Prasad G. Efficacy of medicinal plant (*Andrographis paniculata*) extracts on aflatoxin production and growth of *Aspergillus flavus*. *Lett. Appl. Microbial.* **15** 131.
- Satya S, Reeta Vijayarani. K, Srividhya, R, Gangatharan. N, Francistavier M, Arunprasad, S and Puratchikody A 2003, Antimicrobial screening of leaves of *Memecylon umbeltatum*. Ancient science of life. XX111(2) 120-122.
- 6. Abubaker M N and Ramanathan R 2003, Efficacy of *Euphorbia splendens* and *Leonotis neptaefolia* on aflatoxin producing fungi *Aspergillus flavus* and *A. parasiticus. Indian J. Exp. Biol.* **41** 1473-1475.
- Natarajan E and Francis Xavier T 2003, Sensitivity of Aspergillus species to Tinospora cordifolia (Willd.) Hook.f. and Thomson leaf extracts. Asian J. Microbiol.Biotech. Envi.Sci. 5(4) 543-545.
- 8. Alade, P.I. and Irobi, O.N. 1993. Antimicrobial activities of crude leaf extracts of *Acalypha wilkesiana*. *Journal* of Ehtnopharmacology **39** 171-174.
- Irobi O N and Adedayo O 1999, Antifungal activity of aqueous extracts of dormant fruits of *Hyphaene thebaicia* (Palmae). *Pharmaceutical Biology* 37(2) 114-117.