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# MICROPROPAGATION OF VAJRADANTI (*BARLERIA PRIONITIS* L.): A HIGH VALUE ANTIDONTALGIC AND ETHNO-MEDICINAL HERB

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Tissue culture of medicinal plants is widely used to produce clones and has been found in the conservation of threatened plants species face risk of extinction. Vajradanti is an ethno-medicinally important plant showing pharmacological effects such as anti-microbial, anti-helminthic, anti-fertility, anti-oxidant, anti-diabetic, anti-arthritic, hepato-protective, diuretic, cytoprotective, antidiarrhoeal, analgesic, anti-leukemic, anti-inflammatory and hypoglycemic properties. Micropropagation in most of the plant species involve proliferation of explants using MS medium. There are scanty reports on tissue culture and regeneration of Vajradanti. However, no effective *in vitro* regeneration protocol has been developed for this medicinally important plant. Development of high frequency regeneration pathway was undertaken using shoot tip and nodal segments of *Barleria prionitis* on MS medium supplemented with auxin and cytokinins alone and in various combinations.

Keywords: Conservation; Micropropagation; Regeneration; Vajradanti.

## Introduction

Barleria prionitis (L.) is an ethnomedicinally important herb commonly called Porcupine flower, belonging to family Acanthaceae. Barleria is а large. widespread, polymorphic, pan tropical genus comprised of approx. 300 species of herbs and shrubs. Because of its anti-dontalgic properties it is also known as 'Vajradanti'<sup>1</sup>. Whole plant of *B. prionitis* is used in many ayurvedic formulations and preparations like Rasnadi Kvatha, Rasnadi Churna, Sahachara Ghritha, Sahachara Taila and Dantaroganashani Churna<sup>1,2</sup>. People of many tribals of India-Bheel and Garasia, chew the leaves to relive body ache, toothache and to cure bleeding gums; they

use dried powder of flowers with sugar to make a paste called *Gulkand* which is taken orally for few days to cure painful menstruation. B. prionitis is widely distributed throughout Africa, India, Sri Lanka and tropical Asia<sup>3</sup>. It is planted as an ornamental and cultivated plant in Asia<sup>4</sup>. Barleria is considered as an endangered species<sup>5-7</sup>. Barleria possesses undoubtedly a number of medicinal properties and widely used in treatment of gastro-intestinal disorders, constipation, intestinal worms, digestive troubles, liver disorders, whooping cough, toothache<sup>4</sup>, respiratory syncytial and joint pain<sup>8</sup>. The International Union for Conservation of Nature (IUCN) Species survival commission report "Extinction

crisis continues apace'' for the year 2009 revealed that 70% of plants are under threat. This new era of medicine world immediate rising demand of plant-based drugs is unfortunately creating a pressure on some important medicinal plant populations. High demand and over-exploitation of this species by local ayurvedic practitioners, this plant species is likely to be under threat very soon.

Plant tissue culture technique can provide an alternate source of ex situ conservation and large-scale production of many medicinally important and endangered species<sup>9,10</sup>. In recent years, uses of plant growth regulators in in vitro techniques have been found increasing for conservation of high value plants. Barleria exhibits very low percentage of seed viability<sup>11</sup>. In recent years, many workers have made efforts to micropropagation develop in vitro protocols<sup>7</sup>. MS<sup>12</sup> medium is generally used for the plant propagation through shoot tip and leaf explants on various concentrations of plant growth regulators. A protocol for callus induction has been reported by<sup>12-14</sup>. a detailed study has In a review. been provided regarding the fast depletion of plant resources and their present status<sup>18</sup>.

The present investigation deals with shoot induction and vegetative propagation of *Vajradanti* under *in vitro* conditions, and exploring the effect of different plant growth regulators.

Shot tips were inoculated on MS medium supplemented with BAP, Kn and coconut water in various concentrations and combinations. Maximum (96%) shoot induction or bud break was obtained on MS medium supplemented with 8.88  $\mu$ M BAP giving an average of 3.82±0.13 shoots per explant and shoot length approx 1.74±0.08 cm. The elongated shoots were excised from

mother explant and further multiplied on MS medium supplemented with 4.44  $\mu$ M BAP. **Material and methods** 

Collection of plant material and surface sterilization: Wild plants of Barleria, growing in the forest of Shahabad town near Baran district of Rajasthan were identified as source plants for experimentation. Plants were also grown in the earthen pots so as to monitor the growth and to collect plant material for further experiments.

Healthy shoot cuttings were excised from 4-5 year old plant. After excision, the plant material was washed under running tap water for 10-15 min and then washed with 0.1% Labolene (Qualigens-Fisher) mild detergent. Plant material was also subjected to treatment with antifungal and antibacterial agent (Bacitracin; Hi Media). The plant material was then surface sterilized with 70 % ethanol for 30 sec. and again rinsed twice with double distilled water. Further sterilization of plant material was carried out by 0.1% HgCl<sub>2</sub>, for 3 min. and washed 5 times with autoclaved distilled water under laminar air flow hood. The medium containing 3% sucrose was solidified with 0.8-1 % agar (Hi Media). pH of the nutrient medium was adjusted to 5.8±0.2. Medium was sterilized bv autoclaving at 121°C and 15 psi pressure for 15-20 minutes after dispensing in 100 ml Erlenmeyer flasks and plugged with non cotton. absorbent sterile Shoot tips containing shoot apex (1.0-1.5 mm) were transferred in culture tubes containing MS medium supplemented with 6-benzyl aminopurine (BAP), Kinetin (Kn) and anaphthaleneacetic acid (NAA) alone or in combinations.

All culture vessels were incubated at temperature  $26\pm2^{\circ}C$  with a relative humidity of 50-60% and a photoperiod of 16h per day at 2000-2500 lux provided by fluorescent

incandescent tubes. For each experiments a minimum 7 replicates were taken and repeated thrice.

The nodal segments were also cultured on nutrient medium supplemented with BAP, Kn and NAA in the same manner

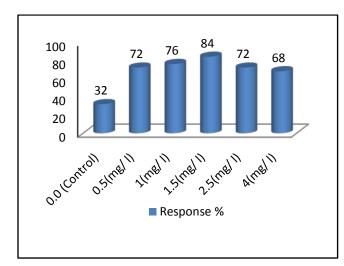
as described above for shoot tips. **Results and Discussion** 

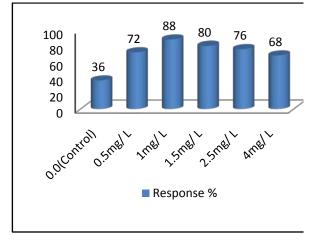
In the recent years, very expressive tissue culture and micro-propagation studies have been carried out in various plant species of family Acanthaceae.

**Table 1**. Response of bud break from cultured shoot tip of Vajradanti (*Barleria prionitis* L.) on

 MS medium supplemented with BAP

Hormone Concentration BAP/ Kn (mg/ L)	No. of explants cultured	No. of explants responded	Response type				
BAP							
0.0 (Control)	25	9	+				
0.5	25	18	+++				
1.0	25	19	+++				
1.5	25	21	++++				
2.5	25	18	+++				
4.0	25	17	+++				
Kn							
0.5	25	18	+++				
1.0	25	22	++++				
1.5	25	20	+++				
2.5	25	19	+++				
4.0	25	17	+++				





**Fig.1.** A *Barleria prionitis*; graph showing percentage response of shoot tip cultured on MS medium supplemented with BAP

**Fig.1. B** *Barleria prionitis*; graph showing percentage response of shoot tip cultured on MS medium supplemented with Kn

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Hormone (mg/		No. of explants cultured	No. of explants responded	Response type	Response (%)
BAP	NAA				
0.0	0.0	25	9	+	36
1.5	0.0	25	3	+	12
1.5	0.5	25	18	+++	72
1.5	1.0	25	24	++++	96
1.5	1.5	25	20	+++	80
1.5	2.0	25	19	+++	76
1.5	2.5	25	16	+++	64
Kn	NAA				
1.0	0.0	25	12	+++	48
1.0	0.5	25	23	+++	92
1.0	1.0	25	19	++++	76
1.0	1.5	25	20	+++	80
1.0	2.0	25	19	+++	76
1.0	2.5	25	13	+++	68

BAP- Benzyl amino purine; Kn- Kinetin; NAA-  $\alpha$  Nephthalene Acetic Acid ; (M±SE)= Mean± Standard Error

#### **Response Type-**

81< ++++ Excellent ; 61-80 +++ 25-40 + Poor ; >25 - Best ; 41-60 ++ Good Negligible

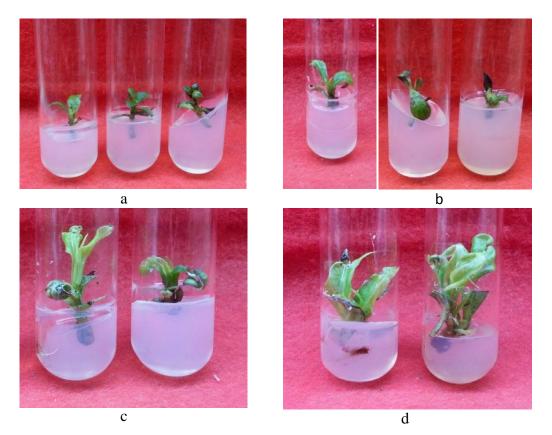
A few prominent among them are, Adhatoda vasica<sup>16</sup>, Dipteracanthus prostates (Robert et al. 2012)<sup>17</sup>. Preliminary research on *in vitro* callus production in *B. prionitis* L. has been reported<sup>13,14</sup>.

*Effect of Cytokinins (BAP and Kinetin:* In the Preliminary experiments, the explants responded to BAP better than other cytokines, hence BAP was used. The fungal infection rate of explants varied from 25-40%. A period of surface sterilization exceeding to 5 minutes was found to be lethal to explants. To protect this infection the spray of antifungal agent was done on the explants after inoculation onto MS medium. Earlier, callus induction response was reported in *B. cristata* on basal defined

nutrient medium supplemented with Kn and dicamba<sup>15</sup>.

After 2 weeks induction response in explants (shoot bud or nodal shoots) was observed on the medium added with BAP alone (1.5mg/L) and Kn (1mg/L), (Table-1; Fig.-1A ,1B). Earlier, a high frequency micropropagation response was observed in *Barleria* using TDZ besides BAP and various auxins. The explants used in this study were Apical, axillary shoots and nodal segments<sup>19</sup>.

*Effect of BAP, Kinetin combination with NAA:* Bud break in explants (shoot tip or nodal segments) were found to be maximum on medium fortified with BAP (1.5) and NAA (1mg/L). Similarly, Kn was also tried in combination with NAA in various J. Phytol. Res. 29 (1 & 2): 1-6, 2016



**Fig.2**. Effect of various hormones on *in vitro* bud breaking from (a) Bud break on MS medium supplemented with BAP (b) Bud break on MS media supplemented with Kinetin (c) Bud break on MS media supplemented with BAP (d) Bud break on MS medium supplemented with Kinetin.

concentrations. It has been found that Kn This best concentration took as further in the combinations of hormones and found maximum response in *Barleria*.

## REFERENCES

- Bhogaonkar PY and Lande SK 1. 2012. Anatomical Characterization of Barleria prionitis Linn.: Α Well known Medicinal herb. Biologia Forum-an internal journal. 4 1-5.
- Sharma PC, Yelne MB, Dennis TJ and Joshi A 2002, Database on Medicinal Plants Used in Ayurveda & Siddha. Central Council for Research in Ayurveda & Siddha, Deptt. of ISM & H, Min. of Health & Family Welfare, Government of India.

- Chavan CB, Shinde UV, Hogade M and Bhinge S 2010, Screening of *invitro* antibacterial assay of *Barleriaprionitis* Lin. J. Herbal Med Toxicol. 4(2) 197-200.
- 4. Burkill HM 1985, *The Useful Plants of West Tropical Africa* Royal Botanic Garden, Kew, UK) **6** 960.
- 5. Khan TI, Dular KA and Solomon MD 2003, Biodiversity conservation in the Thar desert; with emphasis on endemic and medicinal plants. *The Environmentalist.* **23**137-144.
- Pandey R. P., Meena S. L., Pandey P. M., Singhadiya M. K., (2012), Review of depleting plant resource, their present status and conservation in

Rajasthan, India. *Biological Forum*, **4**(1) 213-230.

- Lone SA, Yadhav AS, Bajaj A, Sharma AK, Badkhane Y and Raghuwanshi DK 2013, Conservation6 strategies for threatened medicinal plant – Barleria prionitis L. –using in vitro and ex vitro propagation techniques. Archives of Phytopathology and Plant Protection. 45(11) 1327-1340.
- Parrotta JA 2001, Healing Plants of Peninsular India (CABI Publishing. Wellington, UK & New York) 917.
- Singh SK, Rai MK, Sahoo L 2012, An improved and efficient micropropagation of *Eclipta alba* through transverse thin cell layer culture and assessment of clonal fidelity using RAPD analysis. *Industrial Crops and Products.* 37 328-333.
- 10. Thyagarajan M, Venkatachalam P (2012), Large scale in vitro propagation of Stevia rebaudiana (Bert) for commercial application: pharmaceutically important and antidiabetic medicinal herb. Industrial Crops and Products. 37 111-117.
- 11. Menges ES and Gordon DR 1996, Three levels of monitoring intensity for rare plant species. *Nat. Areas J.* **16** (3) 227-237.
- 12. Murashige T, Skoog F 1962, A revised medium for rapid growth and bioassays with tobacco tissue cultures. *Physiol. Plant.* **15** 473–479.
- 13. Premjet D, Premjet S, Arthur R, Lelono A and Tachibana S 2010, Callus induction and determination of

iridoid glycosides from Barleria prionitis Linn. leaf explants. *Australian Journal of Basic and Applied Sciences.* **4** 4461–4467.

- 14. Shukla P, Singh A, Gawri S, Alexander A and Sonwane S 2011, In vitro propagation of Barleria prionitis L. and its antibacterial activity. International Journal of Pharma Professional's Research. 2(1) 198– 200.
- Abd-El-Mawla AMA, Ahmed AS, Ibraheim ZZ and Ernst L 2005, Phenylethanoid glycosides from *Barleria cristata* L. Callus cultures. *Bull. Pharm. Sci., Assiut University*, 28 (2) 199-204.
- Abhyankar G and Reddy VD 2007, Rapid micropropagation via axillary bud proliferation of *Adhatoda vasica* Ness from nodal segment.*Indian Journal of Experimental Biology*. 45 (3)268-271.
- Robert J, Ravi BX and Louis C 2012, An efficient *in vitro* plant regeneration of *Dipteracanthus prostrates* (Poir.) Ness – a medicinal herb. *Asian Pacific Journal of Tropical Biomedicine*. 484-487.
- Pandey RP, Meena SL, Pandey PM and Singhadiya MK 2012, Review of depleting plant resource, their present status and conservation in Rajasthan, India. *Biological Forum.* 4(1):213-230.
- Lone SA, Yogesh B, Sharma AK, Bakshi AK and Raghuvanshi DK, 2011, Effect of different plant growth regulators on in-vitro propagation of *Barleria prionitis* L. – a threatened medicinal plants. *Int J Pharma and Biosciences*. 2 (1) 438-444.