

ANTISPERMATOGENIC EFFECT OF BENZENE EXTRACT OF *PLUMERIA BICOLOR* LEAVES IN MALE RATS

G. C. JAIN

Department of Zoology, University of Rajasthan, Jaipur - 302004, India.

The effect of benzene extract of *Plumeria bicolor* leaves was studied on testicular function in male rats. Administration of extract (500 mg/kg b.wt./day/orally) for 60 days caused a significant decline in the weights of testes ($P<0.001$), epididymides ($P<0.001$), seminal vesicle ($P<0.001$) and ventral prostate ($P<0.01$) of rats. The sperm counts and motility of cauda epididymal spermatozoa were significantly ($P<0.001$) reduced. The histomorphological picture of the testis of extract treated rats exhibited degeneration of seminiferous epithelium and impairment of spermatogenesis. Leydig cells showed atrophic changes. Thus, the benzene extract of *Plumeria bicolor* leaves has suppressive influence on spermatogenesis and testosterone biosynthesis in rats.

Keywords : Antispermatogetic; *Plumeria bicolor*; Sperm count; Sperm motility.

Introduction

Fertility regulation with natural products of plant origin has been reported in the indigenous system of medicine and appears to be an attractive alternative for a number of reasons. *Plumeria bicolor* (Family Apocynaceae) is a widely grown ornamental plant in Rajasthan, India. Traditionally many species of genus *Plumeria* are used to treat a large number of ailments. The benzene extract of *P. bicolor* leaves possesses anti-implantation effect in female rats¹. Daily feeding of fresh green leaves of *P. alba* to male rats has been reported to cause atrophic changes in testis and arrest of spermatogenesis at primary spermatocyte stage². Phytochemical analysis of the stem bark of *P. bicolor* has shown the presence of ferulic acid derivatives, plumericin and isoplumericin³. In the present study effects of benzene extract of *P. bicolor* leaves on testicular functions of rats were evaluated.

Materials and Methods

Plant material : *P. bicolor* leaves were collected fresh from the University Campus, Jaipur, India and dried under shade. The plant was identified and authenticated at the Herbarium of Botany Department, University of Rajasthan, Jaipur.

Preparation of extract : The shade dried leaves were extracted exhaustively with benzene in the ratio 1:10 (w/v) for 36 hours. The extract was concentrated under reduced pressure and low temperature and was used for the study by suspending in normal saline.

Animals and experimental protocol : Adult colony bred, healthy, male Wistar rats of proven fertility, weighing 220 - 260 g were used for this study. The animals were maintained under controlled temperature ($22 \pm 3^\circ\text{C}$) and constant photoperiodic conditions (12 h light/dark cycle). Commercial

rat feed pellets (Lipton India, Ltd.) and water were provided *ad libitum*. The rats were randomly divided in to two groups of 7 each and treated as follows :

Group I : Vehicle (0.5 ml saline, p.o.) treated control.

Group II : Rats treated with extract (500 mg/kg b.wt./day, p.o.) suspended in normal saline (0.5 ml/rat) for 60 days.

Autopsy : After 60 days of extract treatment the rats were weighed and autopsied under mild ether anesthesia. The testes, epididymides, seminal vesicle and ventral prostate were dissected out, freed from adherent tissues and blood, and weighed. The testes of each animal were fixed in Bouin's fluid for histopathological study.

Sperm motility and density : Cauda epididymal sperm motility and count were determined according to the method of Prasad *et al*⁴.

Testicular histology : Histological sections of testis were cut at 5 μm , stained with Harris haematoxylin and eosin and observed under light microscope for histopathological effects.

Statistical analysis : All the data are expressed as mean \pm SEM. Results were analyzed by Student 't' test. Values of $P<0.05$ were considered as significant.

Results and Discussion

Body and organ weights : The body weight of the control and experimental rats remained significantly unchanged while the relative weights of testes ($P<0.001$), epididymides ($P<0.001$), seminal vesicle ($P<0.001$) and ventral prostate ($P<0.01$) declined significantly (Table 1).

Sperm density and motility : A significant ($P<0.001$) decrease in sperm motility and count was observed in the cauda epididymis of extract treated rats as compared to control rats (Table 2).

Table 1. Body and organ weights of male rats following oral administration of benzene extract of *P. bicolor* leaves.

Treatments	Body weight (g)		Organ weights (mg/100g b.wt.)			
	Initial	Final	Testes	Epididymides	Seminal vesicle	Ventral prostate
Control	230.60±	242.30±	1320±	470.2±	504.5±	481.5±
(vehicle treated)	12.80	18.40	30.8	12.5	32.2	21.7
<i>P. bicolor</i> extract	238.40±	229.50±	1080±	378.4±	316.2±	348.7±
(500 mg/kg b.wt./day)	16.60	17.20	32.5 ^b	16.3 ^b	26.8 ^b	26.8 ^a

Levels of significance

^aP<0.01, ^bP<0.001 compared with control rats.

(Values are mean ± SEM)

Table 2. Effect of benzene extract of *P. bicolor* leaves on histomorphometric characteristics of testis, epididymal sperm count and motility of rats.

Treatments	Number of Seminiferous tubules (%)		Seminiferous tubule diameter (µm)	Leydig cell nuclear diameter (µm)	Sperm count (million/ml)	Sperm motility (%)
	Normal	Abnormal				
Control	89.44±	10.55±	305.05±	6.52±	4.62±	74.30±
(vehicle treated)	2.52	2.05	9.03	0.18	0.17	3.75
<i>P. bicolor</i> extract	41.68±	58.31±	236.57±	4.26±	2.84±	41.42±
(500 mg/kg b.wt./day)	4.05 ^a	4.05 ^a	11.39 ^a	0.17 ^a	0.10 ^a	4.10 ^a

Levels of significance

^aP<0.001

(Values are mean ± SEM)

Histomorphology of testis: Histomorphological study of the testis of vehicle treated control rats showed normal spermatogenesis exhibiting presence of almost all germ cell types in the seminiferous epithelium and normal Leydig cells. In extract treated rats, testicular cytoarchitecture revealed degeneration and impairment of spermatogenesis. The diameters of Leydig cell nuclei and seminiferous tubules were reduced ($P<0.001$) when compared to control. There was significant increase in the number of abnormal seminiferous tubules exhibiting exfoliation and disorganization of cellular stages. The number of round and elongated spermatids and spermatozoa was also declined in affected tubules. The Leydig cells in testis of extract treated rats were atrophic (Fig. 1a, b, Table 2).

The results of the present study shows that benzene extract of *P. bicolor* leaves causes decline in epididymal sperm count and motility and impairment of spermatogenesis in seminiferous tubules.

The weights and functions of sex accessory organs is dependent on testosterone concentration. A significant decrease in the relative weights of epididymides, seminal vesicle and ventral prostate in extract treated rats indicates a suppressed level of testosterone⁵.

Sperm count has been considered to be an important parameter for assessment of male fertility⁶. The depletion in sperm count in cauda epididymis reflects diminished spermatogenesis⁷.

It is well established that the structural and

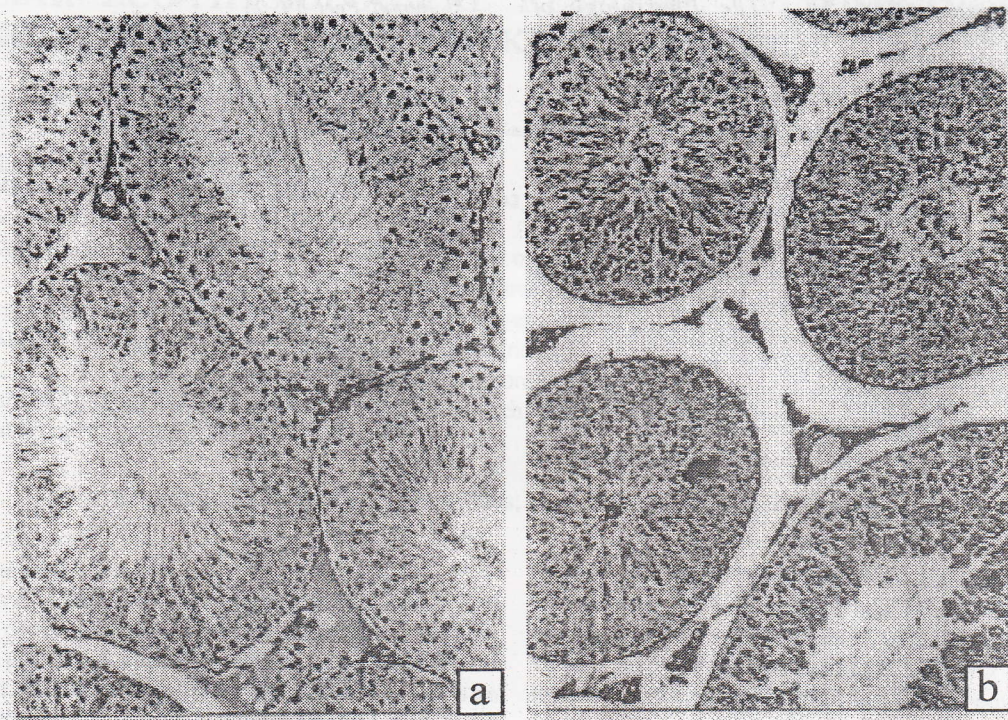


Fig. 1. Photomicrograph of testis

(a) Control rat showing normal spermatogenesis and Leydig cell morphology.

(b) *P. bicolor* extract treated rat exhibiting degenerative changes, impairment of spermatogenesis and atrophy of Leydig cells (X 100, H. E.).

functional integrity of the epididymis are dependent on androgens⁸. Significant decline in the sperm motility in cauda epididymis suggest an impairment of the androgen supply in the epididymis that adversely effects epididymal milieu and consequently sperm maturation, motility and fertilizing ability⁹. Further, the loss of motility of spermatozoa might also be associated with impairment of energy metabolism of spermatozoa¹⁰.

The histological picture of the testis showed degenerative changes, depletion of postmeiotic germ cells. As the spermatogenesis is dependent on testosterone level via its action on the Sertoli cells^{11,12}, the deficiency of testosterone synthesis and secretion is expected to interfere with completion of meiosis resulting in decline in the number of spermatids and spermatozoa. A significant decline in the seminiferous tubule diameter observed in extract treated rats is very likely associated with the decline in the spermatogenic elements and spermatozoa¹³. This is also confirmed by reduced testis weight¹⁴.

The Leydig cells are the most important source of testosterone in the mammalian testis. The decrease in nuclear diameter and atrophy of Leydig cells in extract treated rats suggests impairment of steroidogenesis¹⁵.

The results of the present study indicates that benzene extract of *P. bicolor* leaves shows antispermatogenic effect possibly by testosterone deficiency.

Acknowledgement

The author is grateful to Prof. P. P. Bakre, Head of the Department and Prof. N. K. Lohiya, Coordinator, SAP-UGC for providing necessary facilities.

References

1. Jain GC, Gupta UC and Mathur U 2002, Anti-implantation effect of *Plumeria bicolor* and *Kigelia pinnata* extracts in female rats. *J. Phytol. Res.* 15 41
2. Yadav L 1982, Evaluation of the antifertility effect of some substances of non-steroidal origin and their mode of action in the male rat. Ph.D. Thesis, University of Rajasthan, Jaipur
3. Dobhal MP, Hasan AM, Sharma MC and Joshi BC 1999, Ferulic acid esters from *Plumeria bicolor*. *Phytochemistry* 51 319
4. Prasad MRN, Chinoy NJ and Kadam KM 1972, Changes in succinic dehydrogenase levels in the rat epididymis under normal and altered physiological conditions. *Fertil. Steril.* 23 186
5. Coffey DS 1988, Androgen action and the sex

- accessory tissues. In : *The Physiology of Reproduction*, Knobil E and Neil JD (eds). Raven Press, New York, p. 1081
6. David G, Jouannet P, Martin - Boyce A, Spire A and Schwartz D 1979, Sperm counts in fertile and infertile men. *Fertil. Steril.* **31** 453
 7. Gosh D, Das S, Maiti R, Jana D, Das U 2002, Testicular toxicity in sodium fluoride treated rats : association with oxidative stress. *Reprod. Toxicol.* **16** 385
 8. Brooks DE 1981, Metabolic activity in the epididymis and its regulation by androgens. *Physiol. Rev.* **61** 515
 9. Moore HDM 1998, Contribution of epididymal factors to sperm maturation and storage. *Andrologia* **30** 233
 10. Ford WCL and Harrison A 1987, Futile substrate cycles in the glycolytic pathway of boar and that rat spermatozoa and the effect of α -chlorohydrin. *J. Reprod. Fert.* **79** 21
 11. Steinberger EC 1971, Hormonal control of mammalian spermatogenesis. *Physiol. Rev.* **51** 1
 12. Verhoeven G 1992, Local control systems with in the testis, In : Tindall B (ed.), *Baillier's Clinical Endocrinology and Metabolism*, Vol 6, No. 2, p. 313
 13. Takihara H, Cosentino MJ, Sakatoku J, Cockett, Significance of testicular size measurement in Andrology, 11. Correlation of testicular size with testicular function *J. Urol.* **137** 416
 14. Purohit A and Bhagat M 2004, Contraceptive effect of *Curcuma longa* (L.) in male albino rat. *Asian J. Andrology* **6** 71
 15. Ewing LL and Zirkin BR 1983, Leydig Cell structure and steroidogenic function. *Recent Prog. Horm. Res.* **39** 599