FUNGI ASSOCIATED WITH SEEDS OF PEARL MILLET (PENNISETUM GLAUCUM L. BR.) GROWN IN RAJASTHAN AND THEIR PHYTOPATHOLOGICAL EFFECTS

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One hundred twenty six seed samples of Pearl millet (*Pennisetum glaucum* L. Br.) collected from 14 districts of Rajasthan revealed 45 fungal species of 25 genera in addition to Alternaria teuissima, Alternata alternata, Aspergillus candidus, A. flavus, A. fumigatus, Curvularia lunata, C. penniseti, Drechslera halodes, D. tetramera, D. rostrata, Bipolaris setariae, Helminthosporium spp, Fusarium moniliformae, Fusarium oxysporum, Rhizopus nigricans, Trichothecium roseum, which were dominant and affected seed germination, seedling vigour and seedling diseases.

Keywords: Pearl millet; Phytopathological effects; Seed borne fungi; Seedling diseases.

Introduction

Pearl millet, Pennisetum glaucum (L). Br., is the principal food cereal grown on 25 million-hectare of the drought prone semi- arid regions of African and the Indian subcontinent. Pearl millet is the staple diet of underdeveloped tropical and subtropical countries. The total area of the world under Pearl millet cultivation is about 47 million ha¹, out of which about 25 million ha is in India and Africa alone². Pearl millet occupies a pivotal position among the cereal crops grown in arid zone of Rajasthan. The area under cultivation in India was 9.99 m ha and in Rajasthan it was 4.70 m ha. The seeds are infected and contaminated by many serious seed borne fungi while in field or during seed processing or during storage. Seed borne fungi of some pearl millet cultivars were reported by Konde et al.3, Girisham et al.4 and Mishra and Daradhiyar5. Ahmed and Reddy⁶ and Ingle and Raut⁷ also reported some seed borne fungi on pearl millet. Since no study gives systematic and comprehensive data on fungi associated with Pearl millet seeds grown in Rajasthan, the present investigation was undertaken.

Materials and Methods

One hundred twenty six seed samples of Pearl millet from Rajasthan were collected during the crop season of 2005-2006 by Standard Blotter Test and Potato Dextrose Agar Method. For Dry Seed Examination four replicates of 100 seed per sample taken at random were studied. Percent incidence of different deformities, and insect damaged seeds were estimated. For incubation both untreated and 2% chlorine pretreated (400 seed/ samples) for 5 min were sown on moistened blotters and incubated for 8 days. Twenty five samples were studied using Potato Dextrose Agar (PDA) plates⁸. The discolored or diseased seeds were also plated separately to identify the specific fungi associated with them. Seed germination, seedling symptoms and incidence of fungi were recorded (Table 1).

Results and Discussion

Dry Seed Examination: Seed samples of Pearl millet collected from 14 districts of Rajasthan revealed both asymptomatic and symptomatic seeds when observed under stereobinocular microscope. Symptomatic seeds showed various kinds of disorders like seeds with black or brown discoloration (0.28-55%), seeds with white mycelial growth (0.25-34.7%), shrivelled seeds (0.25-63.75%), seeds with spots and lesions (0.25-13.25%), seeds with water soaked spots (0.25-6%), and broken and insect damaged seeds (2.5- 25%). Seeds with brown discoloration on incubation yielded Alternaria alternata, A. brassicola, A. tenussima, Bipolaris setariae, Helminthosporium spp, Curvularia spp and Drechslera spp. Seeds with white mycelial growth were associated with Fusarium spp and Trichothecium roseum. Aspergillus candidus, A. flavus, A. fumigatus, A. niger, A. sulpheureus. Cladosporium herbarum, and Rhizopus nigricans were the main causal agent of spotted and lesioned seeds. Shirivilled seeds were associated with Curvularia lunata, C. penniseti and C. pallescens or their mixed infection with other fungi. These seeds were common in samples from all the districts. Seeds with water soaked spots were affected by bacteria. Broken seeds splitted into fractions and insect damaged seeds belonging to all districts generally yielded saprophytic fungi (Table 1).

Incubation tests: A total of 44 fungal species belonging to 22 genera, saprophytic as well as pathogenic, were observed on Pearl millet seeds in Blotter and PDA Tests (Table 2 and Figs. 1-15). The fungi encountered in PDA

test were mostly common to those observed in SBM. Fungal spp recorded in SBM were Actinomycetes (1-10%) in SBM and in PDA (1-3%), Alternaria alternata recorded in untreated (4-61%) and pretreated seeds (2.5-30%) in SBM samples and in PDA (1-48%). Alternaria brassicola (1-2%) recorded in untreated seeds and did not occur in pretreated SBM and PDA test. Alternaria longissima (1-24%) in untreated and 1-19% in pretreated seeds. A. tenussima (1-41%) in untreated and pretreated seeds and 1-21% in PDA. Aspergillus candidus (1-10%) in untreated (1-8%) and pretreated (1-10%) seeds and (1-18%) in PDA. A. flavus 3-44%, 4-15% and 1-29% infection in untreated, pretreated seed in SBM and PDA respectively. A. niger was recorded in untreated (2 - 57%), pretreated (1-24%) in SBM and PDA (1-25%). Bipolaris setaraie was recorded (1-45%) in untreated, (1-40%) in pretreated seeds in SBM and (1- 34%) in PDA. Curvularia lunata occurred in untreated, seeds (1-68.5%), pretreated (1-41%) seeds in SBM and PDA (1-20%). Drecshlera tetramera was isolated in untreated (1-46%), pretreated (1-30%) and

(1-48%) in PDA. Diplosporium fulvum occurred (1-12%) in untreated and (1-6%) in pretreated seeds in SBM and it was not observed in PDA test. Doratomyces stemonitis occurred (1-18%) in both untreated and pretreated seeds. Fusarium moniliformae was isolated in PDA test (1-34%) and (1-8%) in SBM. Helminthosporium tetramera was observed (1-21%) in untreated and (1-18%) in pretreated seeds in SBM and (1-16%) in PDA. Rhizopus nigricans encountered in untreated (1-15%) and pretreated (1-8%) seeds in SBM and in PDA (1- 34%). Torula herbarum encountered in 5 samples from Kota district. It occurred (1-9.6%) and (1-6.8%) in untreated and pretreated seeds in SBM. Trichothecium roseum occurred (1-22%) in both untreated and pretreated seeds, (1- 12%) in PDA. Epicoccum purpurascens showed 1-3% infection of the fungus in untreated seeds only. The other minor fungi that were not listed in Table- 2 were Alternaria chrysanthemi, Alternria dianthicola, Aspergillus nidulans, A. ochraceous, A. sulpheureus, Botrytis cineria, SBM. Although the percent range of Aspergillus flavus and A.

 Table 1. Incidence of various seed disorders in Dry Seed Examinations, microorganisms associated and seedling diseases

 caused by them in Standard Blotter Method.

Types of seed discoloration	Percent range	Important microorganism associated with seed	Seedling symptoms	
Seeds with Black or Brown discoloration	0.28 – 55%	Alternaria alternata, A. tenussima	Browning of radical and hypocotyl and brown necrotic	
		Bipolaris setariae	Brown flecks; fine linear streaks, small oval spots.	
		Curvularia lunata, C.penniseti, Drechslera spp.	Light brown spots and streaks at the margin of coleoptile.	
Seed with white mycelial growth	0.25–34.75%	Fusarium spp.	Spotted or lesioned symptoms.	
		Trichothecium roseum, Rhizopus nigricans	Seed and seedling rot.	
		A.niger, A. flavus, A. fumigatus	Yellowing of leaves and radicle rot, brown to black lesions on seedlings.	
Shrivelled seeds	0.25%- 3.75%	Curvularia lunata, C. penniseti, C. pallecsens	Small yellow brown spots on leaves expand to oblong lesions. Center of lesion changes to brown and margins remain yellow.	
Seeds with spots or lesions	0.25-13.25	Trichothecium roseum, Rhizopus nigricans	Seed and seedlings rot.	
Seed with water soaked spots	0.25-6.0%	Bacteria	Mucilaginous bacterial ooze on cotyledons.	
Broken and Insect damaged seeds	2.5-25.0%	Aspergillus spp.	Browning root and radicle rot.	

Table 2. Number of seed samples of Pearl millet infect	ted with fung	gi and perce	ntage range o	f incidence in	incubation tests
(126seed samples studied).					

	Blotter Test				PDA Test	
	Untreated seeds		Pretreated seeds			
Name of the fungi	Samples	%	Samples	%	Samples	%
	infected	range	infected	range	infected	range
Actinomyces	19	1 -10	11	1 -10	5	1-3.0
Alternaria alternata	78	4-61	56	2-30	48	1-48
Alternaria longissima	25	1-24	20	1-19	11	-
A. tenussima	36	1-41	31	1-41	24	1-21
Aspergillus candidus	32	1-8	29	1-10	12	1-18
A. flavus	66	3-44	55	4-15	29	1-29
A. niger	55	2-57	44	1-24	20	1-25
Bipolaris setariae	31	1-45	24	1-40	17	1-34
Chaetomium globosum	− 21 μ 1 1		•	-	2	2-4
Cladosporium cladosporiodes	18	1-33	16	1-37	12	1-26
Curvularia lunata	117	1-68.5	99	1-41	54	1-20
C. pallescens	13	1-6	10	1-3	6	1-7
C. penniseti	21	1-23	21	1-13	12	1-8.0
Diplococcium spp.	12	1-8	-	- *	-	-,
Diplosporium fulvum	4	1-12	4	1-6	-	-
Doratomyces stemonitis	5	1-18	5	1-18	÷	-
Drechslera tetramera	76	1-46	58	1-30	51	1-48
Drechslera halodes	25	1-14.5	13	1-12	10	1-6
D.rostrata	2	1-2	2	-	3	1-4
Epicoccum purpurascens	2	1-3	-	-	-	, -
Fusarium moniliformae	7	1-8	7	1-8	5	1-34
Fusarium oxysporum	6	1-5	7	1-5	9	1-8
Helminthosporium tetramera	29	1-21	18	1-18	12	1-16
Penicillium spp	22	1-12	9	1-8	14	1-9
Rhizopus nigricans	23	1-15	9	1-8	12	1-34
Stachybotrys parvispora	9	1-4	1	1-2	5	1-8.5
Torula herbarum	7	1-9.6	7	1.6.8	-	- ¹ ,
Trichothecium roseum	10	1-22	9	1-22	10	1-12
Verticillium alboatrum	7	1-5	4	1-6	3	1-7

Cephalosporium sp., Cladosporium oxysporum, Cercospora penneseti, Drechslera demtioidea, Pyricularia grisea, Drechslera avenacea, Paecilomyces sp., Chaetomium sp. Fusarium solani, Fusarium semitectum.

Effect of Sodium Hypochloride Pretreatment on Seed Borne Fungi in SBM: In Standard Blotter Method both untreated and seeds pretreated with sodium hypochlorite were used. In general, 1% concentration of available chlorine was found to increase the seed germination without affecting the incidence of the pathogenic seed borne fungi. But the incidence of saprophytic fungi was greatly reduced and their growth and sporulation on seed surface rendered sparse due to seed treatment. The fungi such as *Alternaria brassicola*, *A. chrysanthemi*, *Drechslera rostrata*, *Epicoccum purpurascens* that occurred in low incidences were completely inhibited after chlorine pretreatment in



Figs. 1-15. Photomicrograph showing fungi associated with seeds of Pearl millet (*Pennisetum glaucum* L. Br.) (45x)
1. Bipolaris sorokinianum; 2. Doratomyces stemonitis; 3. Alternaria spp.; 4. Diplococcium fulvum;
5. Curvularia lunata; 6. Drecshlera rostrata; 7. Drecshlera avenaceum; 8. Trichothecium roseum;
9. Torula herbarum; 10. Drecshlera tertramera; 11. Mammaria echinobotryoides; 12. Stachybotrys spp.;
13. Helminthosporium spp. 14. Fusarium oxysporum; 15. Curvularia penniseti.

niger 3.0-44% to 4-15%, 2.0-57% to 1-24% respectively, that of Alternaria alternata 4.0-61% to 30%, of the total fungi recorded, Alternaria alternata, A. tenussima, Aspergillus flavus, A. niger, Rhizopus nigricans, Curvularia lunata, Drechlera tetramera, Fusarium oxysporum, Helminthosporium tetramera, Trichothecium roseum were also important and showed high incidence. Beside the fungal infection, mucilaginous growth of bacteria (0.2 to 9.2%) was observed in 52 samples.

A general assessment of the total seed borne inoculum revealed that seed samples from Alwar, Jaipur, Kota, Jhalawar, Udaipur and Sawai- madhopur mostly 2. showed heavy inoculum and greater incidence of fungi. This may be due to more reverine areas and with more rainfall, hence high humidity is in general, which favours 3. the sporulation of the fungus.

Phytopathological Effects: Fungi associated with seed affected germination as well as vigour and also produced symptomatic seedlings. In 126 samples studied, the germination ranged from 1- 100% in untreated and pretreated seeds in SBM. The fungi which commonly affected seed germination were spp. of Alternaria, Aspergillus, Bipolaris, Curvularia, Drechslera, Fusarium, Helminthosporium and Trichothecium.

Most of the fungi like Alternaria alternata, Aspergillus flavus, Drechslera tetramera, Fusarium moniliformae, Curvularia lunata, C. penniseti, Bipolaris 6. setariae caused serious seed diseases and produced infected seedlings. Seed infection by Alternaria alternata caused swelling of hypocotyl with brown necrotic streaks and browning of radicle. Seed contaminated with 7. Aspergillus flavus caused brown to black lessions on seedlings. Seed infection by Bipolaris setariae caused brown flecks, fine linear streaks, small oval spots, and large 8. irregular oval, oblong or almost rectangular spots.Navi et al.⁹ also reported Bipolaris panici-miliaces on pearl millet.

Fusarium oxysporum caused yellowing and 9. drying of seedlings. Well¹⁰ reported top rot of Pearl millet caused by *Fusarium moniliformae*. Infection with *Curvularia lunata* and *C. penniseti* caused small yellow 10 brown spots on leaves, expands to oblong lesions, center of lesion change to brown and margins remain yellow. 11 Lesions are more common on leaf margins. Infection of seedlings by *Drechslera tetramera* and *D. rostrata* results 12 in 1 to 3 mm long coalescing lesions with extensive necrosis ^{11,12}.

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References

- Ryan J G and Oppen M V 1984, Global production and demand for Sorghum and Millet to the year 2000. Proceedings of the International Symposium on Agrometeorology of Sorghum and Millet in the semi arid tropics (Eds. Virmani, S. M. and Siva Kumar, M. V. K.) ICRISAT, India.pp 41-62.
- Kumar K A and Andrews D J 1993, Genitics of qualitative traits in Pearl millet: A Review. *Crop Sci.* 23 1-20.
- . Kondo B K, Dhage B V and More B B 1980, Seed Borne Fungi of some Pearlmillet Cultivars. *Seed Res.* 8 59-63.
- Girisham S, Rao G V and Reddy S M 1985, Mycotoxin producing fungi associated with pearl millet (*Pennisetum typhoides* (burm. F.) stapf and C. E. Hubb.) *Nat. Acad. Sci. Lett.* 8 333-335.
- Mishra N K and Daradhiyar S K 1991, Mold flora and Aflatoxin contamination of stored and cooked samples of Pearl millet in the Paharia Tribal Belt of Santhal Pargana, Bihar, India. *App. Enviro. Microbiol.* 57 1223-1226.
- Ahmed K M and Reddy C R 1993, A Pictorial guide to the identification of seed borne fungi of Sorghum, Pearl millet, Chickpea, Pigeonpea and Groundnut. ICRISAT, Patencheru, A. P., India, Information Bulletin No. 34.
- 7. Ingle R W and Raut J G 1994, Effect of Fungicidal Sprays on Incidence of Seed Borne Fungi and germination of Pearlmillet. *Seed Res.* **32** 85-87.
- 8. Anonymous 1985, International rules for seed testing. International Seed Testing Association. *Seed Sci. and Tech.* **13** 299-513.
- Navi S S, King S B and Singh S D 1997, A new report of Bipolaris panici- miliaces on pearl millet. International Sorghum and Millets Newsletter 38 124.
- 10. Well H D 1956, Top rot of Pearl millet caused by *Fusarium* moniliformae. Plant Disease Reporter **40** 387.
- 11. Wilson J P and Hanna W W 1992, Disease Resistance in wild *Pennisetum* sp. *Plant Disease* **76** 1171-1173.
- Shetty H S, Mathur S B, Neerguard P and Safeeula K M 1982, *Drechslera setariae* in Indian_Pearl millet seeds, its seed borne nature, transmission and significance. *Transactions of British Mycological Society* 78 170-173.