ASSESSMENT OF TOXICITY OF *PARTHENIUM HYSTEROPHORUS* LINN. LEAF EXTRACT ON LIVER BIOCHEMISTRY OF *RATTUS RATTUS*

S.K. SINGH and B.K. GUPTA

Department of Zoology, M.S.J. Govt. P.G. College, Bharatpur-321001, Rajasthan, India.

Parthenium hysterophorus Linn., an annual herb popularly known as congress grass, is a member or family Asteraceae. The *Parthenium* leaf extract was orally administered to *Rattus rattus*, at 400 mg/ kg b.wt. for acute and 13.33 mg/kg b.wt. for sub-chronic (thirty days) treatment to study its impact on hepatobiochemical parameters. The leaf extract caused a significant fall in acute, and non significant rise after 3 and 30 days of subchronic intoxication. The levels of cholesterol and total lipids were registered at an elevated level after both the treatments. The observed hepato-biochemical changes are suggestive of liver dysfunction and additional mode of action of *Parthenium* leaf extract.

Keywords : Liver cholesterol; Liver glycogen; Liver total lipid; Parthenium hysterophorus Linn; Rattus rattus.

Parthenium hysterophorus Linn., an annual herb of neotropical origin which now has a pantropical distribution¹, belongs to family Asteraceae. Parthenium is an exotic weed popularly known as congress grass, carrot weed, gajar ghas, ramphool, fever few etc. It has been introduced in India about five decade back in mid fifties² and has spread over an area of about 2,05,000 hectares³. Nowadays it can be easily observed on waste lands, railway lines, highways, marshy lands, pond sites, canal ditches, forest and agricultural land associated with every crop⁴. Parthenium has already been reported as a medical hazard causing allergic contact dermatitis, asthma, bronchitis and hay fever in man and live stock⁵.

Parthenium leaf contains non-alkaloid, nonglycosidic sesquitenpene lactone parthenin (α -methylene- γ -lactone) of pseudoguainolide class⁶. Distribution of parthenin in different parts of *Parthenium hysterophorus* shows that leaf contains maximum amount *i.e.* 3.40 percent of wet weight as compared to other parts of the plant⁷.

Although *Parthenium* has adverse effect on man, cattle and plants, it has also been regarded as beneficial plant. It is used in various afflictions like fever, anemia, hepatic ambiosis, dysentry and purification of blood⁸. *Parthenium* has potential use in compost and green manure value⁹. Therefore, study was under taken to assess the toxicity of *Parthenium* leaf extract on liver biochemistry of *Rattus rattus*.

Plant Material: Plants of *Parthenium* were collected from M.S.J. College Campus, vicinity of Keoladeo National Park and residential colonies of Bharatpur, Rajasthan

(India) in the month of August and September, when there was luxuriant growth of plant after rainy season. The leaves were dried at room temperature for fifteen days. Dried leaves were ground in mixer grinder to make fine powder. The powder was mixed with acetone and extraction was carriedout with the help of a soxhelat apparatus for forty eight hours. A brownish sticky resinous material was obtained. The sub lethal dose were prepared by adding distilled water.

Experimental Animal : Colony of healthy *Rattus rattus* were developed and the animals were maintained in metal cages under controlled temperature $25^{0}\pm 5^{\circ}$ C, relative humidity $65 \pm 5\%$ and photoperiod 12 hours / day. The rats were fed on Gold Mohar rat feed (23.5% protein, 5% fat and 4.5% fiber) purchased from Hindustan Lever Ltd., Calcutta. The water was provided *ad libitum*.

Experimental Protocol: The rats of almost same weight, 225 ± 10 gm, were selected randomly irrespective of age and sex. The *Rattus rattus* were grouped into five sets having six individuals in each set. One set for acute study and four sets for subchronic studies. The control set of six rats was run simultaneously for acute and subchronic treatments.

The doses were selected after determining LD_{50} by the log probit analysis method¹⁰. The LD_{50} was calculated as 451.15 mg/kg. b. wt. through oral route of exposure. The *Parthenium hysterophorus* leaf extract was orally administered to *Rattus rattus* at 400 mg/kg. b. wt. for acute and at 13.33 mg/kg. b. wt. for each subchronic treatment by stomach tube feeding. The vehicle distilled

Singh & Gupta

Treatment	Days	Control			Treated			
		Body Weight Liv		er Weight	Body Weight	Liver Weight		
		Wt (g)	Wt (g)	Liver body wt ratio (g)	Wt (g)	Wt (g)	Liver body wt ratio (g)	
Acute	1	227.10 ± 4.20	6.35 ± 0.35	$\begin{array}{r} 0.018 \\ \pm 1.12 \ge 10^{-3} \end{array}$	215.90 ±4.26 A	7.26 ± 0.27	0.021 ± 2.01 x 10 ⁻³	
Subchronic	3	210.60 ± 5.54	6.00 ± 0.17	0.021 ± 2.05 x 10 ⁻³	201.05 ± 4.48	6.71 ± 0.20	$ \begin{array}{r} A \\ 0.023 \\ \pm 1.95 \times 10^{-3} \end{array} $	
	7	215.15 ± 5.14	6.12 ± 0.25	0.019 ± 1.95 x 10 ⁻³	203.10 ± 5.37	6.77 ± 0.30	$\frac{\text{NS}}{0.022} \pm 1.76 \times 10^{-3}$	
	15	212.00 ± 5.32	6.14 ± 0.21	0.020 ± 1.85 x 10 ⁻³	197.51 ± 3.74	7.02 ± 0.22	NS 0.021 ± 1.98 x 10 ⁻³	
	30	210.20 ± 4.97	6.17 ± 0.25	0.022 ± 1.45 x 10 ⁻³	195.10 ± 3.73	7.54 ± 0.32	$ \frac{\text{NS}}{0.025} \\ \pm 1.68 \times 10^{-3} \\ \text{A} $	

 Table 1. Body weight, liver weight and liver weight / body weight ratio in Parthenium leaf extract oral administration in Rattus rattus.

Values as mean \pm S.E., Six rats in each group

A : Significantly different from controls at P<0.01 level; B : Significantly different from control at P<0.02 level; NS : Non-significant

Treatment	Days	Glycogen (Mg/g)		Cholester	ol (Mg/g)	Total Lipid (Mg/g)	
	- 1. A. A. S.	Control	Treated	Control	Treated	Control	1
Acute	1	15.52 ± 0.76	12.70 C ± 0.91	24.40 ± 1.94	41.61 C ± 1.95	02.12 ± 0.06	Treated 03.31 D ± 0.09
Subchronic	3	15.57 ± 0.90	16.98 * ± 0.79	24.51 ± 2.02	31.13 * ± 2.27	02.72 ± 0.05	03.26 A ± 0.08
	7	15.65 ± 0.80	14.21 * ± 0.76	24.86 ± 2.17	31.90 * ± 2.06	03.14 ± 0.05	03.57 * ± 0.09
	15	15.79 ± 1.01	13.63 * ± 0.93	24.80 ± 2.15	32.81 * ± 2.24	3.15 ± 0.05	03.90 A ± 0.11
	30	15.58 ± 1.12	16.20 * ± 0.87	24.62 ± 2.22	32.31 * ±2.13	02.64 ± 0.15	03.52 C ± 0.08

Table 2. Liver biochemical parameters after Parthenium leaf extract oral administration in Rattus rattus.

Values are mean \pm S.E. of 6 determinations, A : P<0.05, * = Non significant

B : P<0.02, C : P<0.01,

D:P<0.001,

water was given to control set of rats.

The rats were reweighed and excised out after the termination of the experimental period *i.e.* 1, 3, 7, 15 and 30 days. Livers were weighed and processed for biochemical studies. Change in body weight and liver weight to body weight ratio was determined. Liver glycogen, cholesterol and total lipids were estimated by the standard methods¹¹⁻¹³. Data were subjected to statistical evaluation using student's t-test. Statistical significances were used at P<0.01, P<0.02 and P<0.05 levels.

On oral administration of an acute dose of *Parthenium* leaf extract rats developed onset of toxicity within 5-7 minutes, exhibiting hypersalivation and tremors. Subsequently, the sign of maximal severity including convulsions, nofeeding, anorexia, ejection of food with equal amount of water (diarrhoea), pulmonary dysfunction (as revealed by labored breathing) leading to leg weakness (body on half paw) were evident within 15-30 minutes and lasted about three hours. However, the toxicity manifestations exhibited by sub-chronically treated rats were of mild severity and lasted about one hour. These signs are suggestive of both central and peripheral origin¹⁴⁻²³.

A significant reducation in body weight, after acute treatment, has been observed as compared to controls (Table 1). However, increase (significant) in liver body weight ratio after acute and sub-chronic (30 days) treatment was evident (Table 1). A significant fall in glycogen after acute and non-significant rise after days 3 and 30 has been observed (Table 2). The levels of cholesterol and total lipids were registered at an elevated level after both the treatments (Table 2).

Loss in body weight of rats might be due to anorexia, food avoidance and diarrhoea caused by *Parthenium* leaf extract, as has also been observed in buffalo calves¹⁹. The weight of liver in rats was markedly affected. The change may be due in part, to a direct effect of *Parthenium* leaf extract on liver weight and in part to an indirect effect due to general growth depressing effects of the extract. The indirect effect of *Parthenium* leaf extract on growth could cause change in liver weight^{21,23}. Since the body weight decreases with the increase in liver weight, an increase in liver to body weight ratio is evident and gain support²².

The depletion in liver glycogen may be attributed to anorexia, labored breathing and diahhroea as a result of which demand for glucose may increase in treated rats. During such conditions glycogenolysis might have increased to fulfill the glucose requirement of the rats and is in close affirmation to Ahmed *et al.*²². Further, the

reduction in hepatic glycogen can also be due to the lost capacity of liver to mobilise glucose as glycogen following pathophysiological changes (increased liver weight) as is evident in the present investigation and gain support by other studies^{21,23}.

Increase in total lipid and cholesterol level in liver as a result of *Parthenium* leaf extract treatment probably indicate the impaired lipid and cholesterol metabolism. Since, liver is the major organ of biosynthesis of cholesterol and responsible for mobilization and excretion, the high levels of cholesterol and total lipid in the liver of rats show that the process is highly interrupted leading to accumulation of cholesterol and total lipid in liver.

The lipids and cholesterol are carried by lipoproteins, forming very low density lipoproteins (VLDL) from the liver, to different tissues of the body. The elevation in total lipid and cholesterol level may further be attributed to lower level of lipoprotein in the liver of rats after *Parthenium* leaf extract treatment. It is evident that acute and sub-chronic (30 days) treatment is more prominent.

In conclusion, *Parthenium* leaf extract intoxication to rats lowers glycogen and increases cholesterol and total lipid levels in liver.

References

- 1. Evans HC 1997, *Parthenium hysterophorus* a review of its weed status and the possibilities for biological control. *Biocontrol news and information* 18 389-98.
- 2. Rao RS 1956, Parthenium, a new record for India. J. Bomb. Nat. His. Soc. 54 218.
- 3. Gidwani I 1975, *Parthenium*, a new weed in India, *Pans.* 22 280
- 4. Krishnamurthy K 1976, Parthenium weed the problem of present day. Pesticides 10 33.
- 5. More PR, Vadlamudi VP and Qureshi MI 1982, Toxicity of *Parthenium hysterophorus* Linn. In bovines : Changes in some biochemical constituents of blood. *Indian Vet. J.* **59** 515.
- Subbarao PV, Mangla A, Subbarao BS and Prakash KM 1977, Clinical and immunological studies on person exposed to *Parthenium hysterophorus* Linn. *Experimentia* 33(10) 1387.
- Subbarao PV, Mangla A, Subbarao BS and Prakash KM 1976, *Parthenium* – an allergic weed. Proc. Seminar on (*Parthenium* – a positive danger) West End Hotal, Bangalore Sept. 4, 1976, 17-18. Organised by Bangalore international cities relationship organization and Univ. of Agril. Sciences, Bangalore.
 Ramaswami PP 1977, Potential usages of *Parthenium*
- In : Proc. First International conference on

Parthenium Management, University of Agricultural Sciences, Dharwad, 6-8 Oct., 1997, I 77-90.

- 9. Mahadevappa M 1999, *Parthenium* and its management. Publication Center, University of Agricultural Science, Dharwad. India. 46.
- 10. Finney DJ 1971, *Probit analysis*. Cambridge University Press, pp 303.
- 11. Montegomery R 1957, The determination of glycogen. Arch. Bioch. Biophys. 67 378-387.
- Zlatkis A, Zak B and Boyle A 1953, A new method for direct determination of serum cholesterol. J. Lab. Clin. Med. 41 486.
- 13. Folch J, Less M and Stanley CHS 1957, A simple method for isolation and purification of total lipids from animal tissues. J. Biol. Chem. 226 497.
- 14. Dollahite JW, Hardy WT and Henson JB 1964, Toxicity of *Helenium microcephalum* (Small head Sneezy weed). J. Am. Vet. Med. Ass. 145 694.
- Lonker A, Mitchell JC and Calman CB 1974, Contact dermatitis from *Parthenium hysterophorus*. *Transcripts* of St. John's Dermatological Society 60 43-53
- 16. Narsimhan TR, Anant M, Narayan Swami, Rajendra Babu M, Mangla A and Subbarao PV 1977, Toxicity of *Parthenium hysterophorus* Linn to cattle and buffaloes. *Vet. Bull.* 48 249
- 17. Qureshi MI, Vadlamudi VP and Wagh KR 1980, A

study on subacute toxicity of Parthenium hysterophorus Linn in goats. Live stock Adv. 5 39.

- Hariharan NV, Nandkumar NV and Adinarayana Murthy NV 1982, Effect of *Parthenium* hysterophorus acetone extract on the sciatic nervegasteronemius muscle preparation of frog *Rana* hexadactyla. J. Environ. Biol. 3(3) 103-111.
- Kadhana DL, Jangde CR, Sadekar RD and Joshirao MK 1992, *Parthenium* toxicity in buffalo calves. J. Soils and Crops 2(1) 69-71.
- Vijaylakshmi P, Vijaylakshmi KM and Nandkumar NV 1999, Depolarizing neuromuscular Junctional blocking action of *Parthenium hysterophorus* leaf extract in Rat. *Phytother. Res.* 13 367-370.
- 21. Singh SK and Gupta BK 2006, *Parthenium* hysterophorus Linn. Induced clinical manifestations in *Rattus rattus. J. Phytol. Res.* **19**(2) 331-332.
- 22. Ahmed MN, Rao PR and Mahender M 1988, Experimental introduction of acute toxicity in buffalo calves by feeding *Parthenium hysterophorus* Linn. *Indian J. Animal Sci.* 58 731-734.
- 23. Singh SK 2007, Toxicological evaluation of the congress grass *Parthenium hysterophorus* Linn. on *Rattus rattus*: A study based on behavioral changes, blood and liver biochemistry. *Ph.D. Thesis, University of Rajasthan, Jaipur*.