

ALTERED THYLAKOID MEMBRANE ORGANIZATION AND PHOTOCHEMICAL FUNCTIONS IN WHEAT THYLAKOID MEMBRANES UNDER THE INFLUENCE OF CADMIUM

Md. AZEEMUL HASAN,¹ A. PHANINATHA SARMA,² G. NAGENDRA BABU² and S. D. S. MURTHY²

¹Department of Biochemistry, Chaitanya Post Graduate College, Warangal, A.P., India.

²Department of Biochemistry, Sri Venkateswara University, Tirupati-517 502, A.P., India.

Electron transport measurements of thylakoid membranes isolated from Cd treated eight day old wheat primary leaves indicated that there is a concentration dependent loss of PS II activity and 50% inhibition was noticed at 0.2 mM CdCl₂. Chlorophyll fluorescence measurements demonstrated the existence of inhibitory site at water oxidation complex through the decrease of fluorescence intensity. Lipid Peroxidation measurements of wheat thylakoid membranes clearly gave an indication that free radical mediated lipid Peroxidation of thylakoid membranes is one of the reason for the loss of PS II activity.

Keywords: Cadmium stress; Electron transport; Lipid peroxidation; Spectral properties; Wheat leaves.

Abbreviations: pBQ; Para benzoquinone; Chl, chlorophyll; PS; Photosystem; MDA, Malondialdehyde.

Introduction

Cadmium is strongly phytotoxic and causes growth inhibition and even plant death by some studies using different plant species^{1, 2} however, the mechanisms involved in Cd toxicity are still not completely understood. The plant growth inhibition induced by Cd was probably correlated to its reduced photosynthetic rate, detrimental effects on chloroplast function, cell division³, chloroplast structure⁴, as well as the water splitting apparatus of photosystem (PS) II and photosynthetic electron transports⁵. Chlorophyll fluorescence is an indicator which gives information about the absorption, distribution and utilization of energy in photosynthesis^{6, 7}. Room temperature Chl a fluorescence is associated with the functioning of PS II. Any inhibition of electron flow to the PS II reaction center lowers the Chl fluorescence yield where as block at acceptor site of PS II increases fluorescence⁸. In addition to this, the over production and rapid accumulation of reactive oxygen species (ROS), which is considered as one of the important mechanisms of heavy metal toxicity, are the early responses of plant to heavy metals^{9, 10}. Therefore, a comparative study has been made regarding the effect of Cd on PS II electron transport activity, Chl fluorescence and lipid peroxidation of thylakoids isolated from wheat primary leaves. Our results indicate that Cd exerts inhibition in PS II catalyzed electron transport activity by causing lipid peroxidation in

thylakoid membranes.

Material and Methods

Wheat (*Triticum vulgare*) seedlings were raised on petri plates under continuous white light (160 μ moles m⁻² S⁻¹) at 25°C. Hoagland solution was supplied at 4 day intervals to the seedlings. Eight day old seedlings were exposed to different concentrations of CdCl₂ (0.1 mM - 0.5 mM) for 24 hr. After the treatment, primary leaves of both control and heavy metal treated seedling were sampled for thylakoid membranes isolation and for the assay of photochemical activities.

The thylakoids were used for measurement of photochemical activities by following the procedure of Sabat *et al.*¹¹ with slight modifications. The assay mixture for whole chain electron transport activity contained 0.5 mM MV (Methyl Viologen) and 1mM sodium azide in three ml of the 25 mM HEPES reaction buffer (pH 7.8). For PS II mediated oxygen evolution, the reaction mixture consisted of 0.5 mM pBQ in three ml reaction buffer. The fluorescence emission spectra was measured by exciting the thylakoid membranes with 440 nm light. The slit width for both excitation and emission was 5 nm. Samples were kept in dark for 5 min before measurement of the spectra. Lipid peroxidation has been measured according to the method of Heath and Packer¹². The Malondialdehyde (MDA) calculations were made by using the extinction coefficient 155 mM⁻¹ cm⁻¹. The amount of MDA was expressed as

Table 1. Effect of Cd on the whole chain electron transport activity of the thylakoids isolated from control and Cd treated wheat primary leaves. Three ml of reaction mixture contains reaction buffer 25mM HEPES-NaOH (pH. 7.5) containing 20 mM NaCl, 0.5mM MV, 1 mM Na-azide and thylakoids equivalent to 40 μ g of Chl. Other details were given in material and methods. The SD is not more than 10%.

Concentration of CdCl ₂ , mM	Whole chain electron transport activity H ₂ O → MV μ moles of O ₂ ↓ mg ⁻¹ Chl h ⁻¹	Percentage loss
Control	195 ± 19	0
0.1	141 ± 13	28
0.2	102 ± 9	48
0.3	65 ± 7	67
0.4	45 ± 4	77
0.5	35 ± 3	82

Table 3. Effect of Cd on chlorophyll fluorescence emission properties of thylakoid membranes. Three ml of reaction mixture contains reaction buffer 25mM HEPES - NaOH (pH. 7.5) containing 20 mM NaCl and thylakoids equal to 8 μ g of Chl a Slit width for both excitation and emission was 5 nm.

Concentration of CdCl ₂ , mM	Chlorophyll fluorescence (Rel. units)	Percentage loss
Control	72 ± 7	0
0.1	52 ± 4	28
0.2	36 ± 3	50
0.3	29 ± 3	60
0.4	25 ± 2	65
0.5	23 ± 2	68

nmol of MDA per mg protein.

Results and Discussion

Cadmium induced alterations in whole chain Electron transport- Methyl Viologen (MV) is known to accept the electrons from A₀ in photosynthetic electron transport chain¹³. Therefore, whole chain electron transport activity has been measured in thylakoid membranes using MV as terminal electron acceptor (H₂O → MV). Control thylakoids without heavy metal treatment exhibited a high rate of oxygen consumption (195 μ moles O₂ ↓ mg⁻¹ Chl h⁻¹). Increase in the Cd concentration from 0.1 mM to 0.5 mM brought enhancement in the inhibition of whole chain electron transport. Almost 50% loss was noticed above 0.2 mM of Cd treatment (Table-1). The reason for the loss of whole chain electron transport could be either

Table 2. Effect of Cd on the photosystem II catalyzed electron transport activity of the thylakoids isolated from control and Cd treated wheat primary leaves. Reaction mixture (3 ml) for this assay contained reaction buffer, 0.5 mM pBQ and thylakoid equivalent to 40 μ g of Chl. Other details were given in material and methods. The SD is not more than 10%.

Concentration of CdCl ₂ , mM	Whole chain electron transport activity H ₂ O → pBQ μ moles of O ₂ ↑ mg ⁻¹ Chl h ⁻¹	Percentage loss
Control	271 ± 27	0
0.1	200 ± 19	26
0.2	127 ± 10	53
0.3	91 ± 8	66
0.4	58 ± 6	79
0.5	48 ± 5	82

Table 4. Effect of Cd on lipid Peroxidation of thylakoid membranes of wheat plants.

Concentration of CdCl ₂ , mM	Lipid peroxidation n mol MDA mg protein ⁻¹	Percentage enhancement
Control	42 ± 4.1	0
0.1	50 ± 4.9	19
0.2	62 ± 5.8	47
0.3	75 ± 6.8	78
0.4	78 ± 6.9	85
0.5	74 ± 6.7	75

alterations at the level of PS II or PS I catalyzed electron transport^{13,15,5}. Thus, Cd stress induced inhibition of whole chain electron transport could be due to either alterations at PS II or PS I. To identify the target photosystem, we have measured the partial electron transport reactions mediated by individual photosystems.

Inhibitory effect of Cd on photosystem II catalyzed electron transport - Since Cd inhibited the whole chain electron transport, to find out whether the alterations are due to changes in PS II or PS I, an attempt has been made to study the Cd effect on PS II catalyzed p-benzoquinone (PBQ) supported Hill reaction (Table-2). pBQ is known to accept the electrons from PQ pool¹³. Being lipophilic in nature pBQ can easily enter into thylakoid membrane and reach PQ pool. Control thylakoids exhibited a rate of

oxygen evolution activity ($271 \mu \text{ moles of O}_2 \uparrow \text{mg}^{-1} \text{ Chl h}^{-1}$) Cd treatment caused gradual increase in the inhibitory pattern and maximum loss was observed after giving the treatment with 0.5 mM of CdCl₂. 50% loss was noticed at 0.2 mM of CdCl₂. The reason for the loss of PS II catalyzed electron transport could be either due to alterations at water oxidation complex or due to changes in D₁ and D₂ polypeptides or due to alteration at the level of reducing site of PS II¹⁵⁻¹⁷.

Identification of the target site for Cd action in photosystem II: Chlorophyll fluorescence is an indicator of photo system II photochemistry therefore an attempt has been made to identify the target site in PS II photochemistry using chl fluorescence as a tool (See Table 3). Control thylakoid membranes upon excitation with 440 nm light exhibited a single peak at 685 nm. The fluorescence intensity at this peak position was observed to be 65 rel. units. The increase in the concentration of Cd brought gradual decrease in the fluorescence intensity and 50% decrease was noticed at 0.2 mM concentration of Cd. The decrease in the chl fluorescence clearly indicate the existence of inhibitory site at the donor site of PS II as suggested by Butler⁸. Thus, the reason for the observed inhibition in PS II activity could be due to changes induced by Cd in the water oxidation complex. Since thylakoid membranes are the sites for PS II photochemistry, any change in the thylakoid lipid profile can also lead to the loss of PS II activity. To verify this, a study has been made to study the-lipid peroxidation in control and treated samples.

Effect of Cd on the lipid Peroxidation of thylakoid membranes: Table 4 shows the lipid Peroxidation pattern of control and Cd treated thylakoid membranes. The increase in the Cd concentration from 0.1 to 0.4 mM caused gradual increase in the malondialdehyde (MDA) fractions by 85 %. Further increase in the concentration to 0.5 mM could not cause further enhancement in the lipid peroxidation. In this way, alterations in the thylakoid membrane lipids and changes in the water oxidation complex are together responsible for the altered photochemistry in PS II under Cd stress.

Acknowledgement

We are thankful to Prof. Prasanna Mohanty for making valuable suggestions in the preparation of the manuscript.

References

- Romero-Puertas MC, Palma J M and Gomez M 2002, Cadmium causes the oxidative modification of proteins in pea plants. *Plant, Cell and Env.* **25** 677-686.
- Wojcik M, Vangronsveld J and Tukiendorf A 2005, Cadmium tolerance in *Thlaspi caerulescens* 1. Growth parameters, metal accumulation and phytochelatin synthesis in response to cadmium. *Env. and Exper. Bot.* **53** 151-161.
- Baryala A, Carrier P, Franck F 2001, Leaf chlorosis in oilseed rape plants (*Brassica napus*) grown on cadmium polluted soil: causes and consequences for photosynthesis and growth. *J. Planta.* **212** 696-709.
- Barcelo J, Vazquez MD, Poschenrieder C 1988, Structural and ultra structural disorders in cadmium-treated bush bean plants (*Phaseolus vulgaris* L.). *New Phitol.* **108** 37-49.
- Mallick N and Mohn F R 2003, Use of chlorophyll fluorescence in metal-stress research: a case study with the green microalga. *Scenedesmus Ecoto. and Env. Safety.* **55** 64-69.
- Papageorgiou G C 1975, Chlorophyll fluorescence: An intrinsic probe of photosynthesis. In: *Bioenergetics of Photosynthesis.* (Govindjee, ed.) Pp.320-366, Academic Press, New York.
- Fork D C and Mohanty P 1986, Fluorescence and other characteristics of blue green algae (Cyanobacteria), red algae and cryptomonads. In: *Light Emission by Plants and Bacteria* (Govindjee, Amsez, J. and Fork, D.C., eds.) pp. 451-496, Academic Press, New York.
- Butler WL 1977, Chlorophyll fluorescence as a probe for electron transfer and energy transfer. In: *Photosynthesis I, Encyclopedia of Plant Physiology.* (Trebst, A. and Avron, M., eds.) vol 5, pp.149-167. Springer-Verlag, Berlin.
- Mittler R 2002, Oxidative stress, antioxidants and stress tolerance. *Trends in Plant Sci.* **7** 405-410.
- Hartley-Whitaker J, Ainsworth G and Meharg A A 2001, Copper and arsenate induced oxidative stress in *Holcus lanatus* L. clones with differential sensitivity. *Plant, Cell and Env.* **24** 703-722.
- Sabat S C, Mohanty N and Mohanty P 1986, Heat induced alteration in electron donation sites of ascorbate and ascorbate reduced catechol in the electron transport chain of *Amaranthus* chloroplasts. *Ind. J. Biochem. Biophys.* **23** 266-269.
- Heath R L and Packer L 1968, Photoperoxidation in isolated chloroplast. 1. Stoichiometry of fatty acid peroxidation. *Arch. Biochem. Biophys.* **125** 189-198.
- Trebst A 1974, Energy conservation in photosynthetic electron transport of chloroplasts. *Ann. Rev. Plant Physiol.* **25** 423-458.
- Li E H and Miles C D 1975, Effects of cadmium on photosystem II of chloroplasts. *Plant Sci. Lett.* **5** 33-40.

- 15. Baszynski T, Wajda L, Krol M, Walinska D, Drupa Z and Tukendorf A 1980, Photosynthetic activities of cadmium treated tomato plants. *Physiol. Plant.* **48** 365-370.
- 16. Bazzaz MB and Govindjee 1974, Effects of cadmium

- nitrate on spectral characteristics and light reactions of chloroplasts. *Env. Left.* **6** 175-191.
- 17. Singh DP and Singh SP 1987, Action of heavy metals on Hill activity and O₂ evolution in *Anacystis nidulans*. *Plant Physiol.* **83** 12-14.

new cadmium polluted soil: causes and consequences for plant growth and photosynthesis and growth. *Plant* 212 688-709

4. Barcelo J, Vazquez MD, Poschenrieder C 1988, Structural and ultra structural disorders in cadmium-treated bush bean plants (*Phaseolus vulgaris* L.). *New Phytol.* **108** 37-44

5. Mallick N and Mohan P 2003, Use of chlorophyll fluorescence in metal stress research: a case study with the green microalgae *Scenedesmus Ecoto*. and *Chlorella* sp. *25* 64-69.

6. Papageorgiou G C 1975, Chlorophyll fluorescence: An intrinsic probe of photosynthesis. In: *Bioenergetics of Photosynthesis* (Govindjee, ed) pp 320-366 Academic Press, New York

Lok D C and Mohan P 1986, Fluorescence and other characteristics of pine green algae (*Cyanobacteria*), red algae and cryptomonads. In: *Light Emission by Plants and Bacteria* (Govindjee, ed) pp 421-496 Academic Press, New York

8. Butler W L 1971, Chlorophyll fluorescence as a probe for electron transfer and energy transfer. In: *Photosynthesis II: Encyclopaedia of Plant Physiology* (Teuber A and Avton M, eds) vol 2, pp 149-167 Springer-Verlag Berlin

9. Mittler R 2002, Oxidative stress, antioxidants and stress tolerance. *Trends in Plant Sci.* **7** 402-410

10. Harley-Whitaker J, Ainsworth G and Meharg A A 2001, Copper and arsenite induced oxidative stress in *Holcus lanatus* L. clones with differential sensitivity. *Plant Cell and Env.* **24** 103-112

11. Sapat S C, Mohan N and Mohan P 1986, Heat induced alteration in electron donation sites of ascorbate and ascorbate reduced catechol in the electron transport chain of *Chlorella* sp. *13* 266-269

12. Heim R L and Facker 1968, Photoperoxidation in isolated chloroplasts. I. Stoichiometry of fatty acid peroxidation. *Arch Biochem Biophys* **125** 189-198

13. Hebel A 1974, Energy conservation in photosynthetic electron transport of chloroplasts. *Ann Rev Plant Physiol.* **25** 423-428

14. El-FH and Miles C D 1975, Effects of cadmium on photosystem II of chloroplasts. *Plant Sci Lett.* **5** 3-40

to be 0.2 rel. units. The increase in the concentration of Cd brought gradual decrease in the fluorescence intensity and 50% decrease was noticed at 0.2 mM concentration of Cd. The decrease in the chl fluorescence clearly indicate the existence of inhibitory site at the donor site of P2 II as suggested by Butler. Thus, the reason for the observed inhibition in P2 II activity could be due to changes induced in the water oxidation complex. Since thylakoid membranes are the sites for P2 II photochemistry, any change in the thylakoid lipid profile can also lead to the loss of P2 II activity. To verify this, a study has been made to study the lipid peroxidation in control and treated samples.

Effect of Cd on the lipid Peroxidation of thylakoid membranes: Table 4 shows the lipid Peroxidation pattern of control and Cd treated thylakoid membranes. The increase in the Cd concentration from 0.1 to 0.4 mM caused gradual increase in the malondialdehyde (MDA) fractions. Further increase in the concentration to 0.2 mM could not cause further enhancement in the lipid peroxidation. In this way, alterations in the thylakoid membrane lipids and changes in the water oxidation complex together responsible for the altered photochemistry in P2 II under Cd stress. *Effect of Cd on the lipid Peroxidation of thylakoid membranes: Table 4 shows the lipid Peroxidation pattern of control and Cd treated thylakoid membranes. The increase in the Cd concentration from 0.1 to 0.4 mM caused gradual increase in the malondialdehyde (MDA) fractions. Further increase in the concentration to 0.2 mM could not cause further enhancement in the lipid peroxidation. In this way, alterations in the thylakoid membrane lipids and changes in the water oxidation complex together responsible for the altered photochemistry in P2 II under Cd stress.*

We are thankful to Prof. Prasanna Mohan for making valuable suggestions in the preparation of the manuscript. References are given at the end of the paper in respect of the following authors: Farias M O, Farias M and Gomes M 2002, Cadmium causes the oxidative modification of proteins in *Chlorella* sp. *Plant Cell and Env.* **25** 671-680

San Woidik M, Vengronwid J and Tarkenton A 2002, Lead tolerance in *Chlorella* sp. *Plant Cell and Env.* **25** 671-680