INCIDENCE AND HISTOPATHOLOGY OF ALTERNARIA ALTERNATA IN SUNFLOWER SEEDS

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One hundred eighty eight seed samples of sunflower (*Helianthus annuus L*) revealed asymptomatic and symptomatic (bold-discoloured and shrivelled-discoloured) seeds. Seeds with greyish brown and brown to black discolourations yielded *Alternaria alternata*. The fungus was recorded in 133 seed samples with an incidence range of 1-98%. The pathogen was located in pericarp of asymptomatic seeds whereas, in pericarp, endosperm and embryo of bold-discoloured and shrivelled-discoloured seeds. The mycelium was inter- and intracellular. Seeds with severe infection showed a heavy aggregation of mycelium in the inner layers of the pericarp, space between endosperm and embryo and cells of cotyledons and shoot apex.

Keywords: Alternaria alternata; Histopathology; Incidence; Seeds, Sunflower.

Introduction

The frequent occurrence of Alternaria alternata, internally in cereal seeds is well documented¹⁻³. In oilseeds it has been recorded in linseed⁴, rape and mustard^{5,6} and taramira⁷. A. alternata causes leaf spot in sunflower⁸⁻¹⁰. During routine seed health testing of samples collected from Rajasthan, it was found to occur in large number of seed samples of sunflower; therefore, a study of its incidence in seed lots and location in seeds was made. Materials and Methods

One hundred eighty eight seed samples collected from 11 districts of Rajasthan during 1992-94 were tested by dry seed examination and blotter test¹¹ to study the incidence of *Alternaria alternata*. Location of *A. alternata* was studied in two naturally infected seed samples (ac. nos. 8504 and 8512) carrying 85% and 89% incidence respectively, using methods of component plating, clearing and wholemount preparations and microtome sectioning¹².

Results and Discussion

Incidence in seed samples : Seed samples of sunflower on dry seed examination revealed

bold-symptomless, bold-discoloured and shrivelled-discoloured type seeds (Fig.1). Seeds with greyish-brown and brown to black discolourations on incubation yielded growth of *Alternaria alternata*. In blotter test, 43 fungi of 23 genera were isolated from seeds of which *A. alternata* was recorded in 133 seed samples with incidence range of 1-95% in untreated seeds and 1-98% in pretreated seeds. Samples from Jhalawar, Jaipur, Kota, Baran and Bhilwara revealed relatively high infection percentages.

The heavy infestation of seed samples (70.74%) with *A. alternata* (1-98%) from 11 districts suggests its widespread occurrence in Rajasthan. By producing discolourations on surface it affects the seed quality adversely. Similar observations have also been reported in rape and mustard^{5,6}, coriander¹³ and sunflower¹⁴.

Histopathology of A. alternata infected seeds: In component plating, growth of A. alternata (Fig.2) was recorded on outer- and inner pericarps in bold-symptomless seeds whereas in outer- and inner pericarps, endosperm and embryo in bold-discoloured and

| wholemoule preparate | | | | | | |
|---|-----------------|------------------------------|------------------|----------------|----------------|------------------|
| SEED COMPONENTS | SEED CATEGORIES | | | | | |
| | Ac.no.8504 | | | | Ac.no.8512 | |
| | BS | BD | SD | BS | BD | SD |
| COMPONENT PLATING: | estatio concel. | 11:0 - 20 - 2 - ² | gan e - kez-e a | dyn ordefni | (Jae harden | |
| Seed coat Outer pericarp Inner pericarp | 22 12 00 | 70 66 50 | 100 100 78 | 10 10 00 | 62 52 40 | 100 100 74 |
| Embryo Cotyledons Embryal axis | 00 00 | 36 16 | 60 24 | 00 00 | 28 16 | 40 26 |
| II. CLEARED AND WHOLEMOUNT PF | REPARATION | (S : | | 10 | 40 | 100 |
| Seed coal Outer pericarp Inner pericarp | 14 10 | 60 56 36 | 100 100 68 | 10 10 00 | 40 38 24 | 100 100 72 |
| Endosperm Embryo Cotyledons | 00 00 | 20 10 | 30 18 | 00 00 | 18 10 | 28 18 |

Table 1. Percentage infection of A. alternata in different parts of seeds of sunflower in component plating and cleared 1 _____

BS = Bold-symptomless; BD = Bold-discoloured; SD = Shrivelled-discoloured

shrivelled-discoloured seeds in both the samples (Table 1).

Cleared wholemount preparations revealed thick, dark, branched and septate mycelium of A. alternata in outer pericarp (Fig.3) and inner pericarp in boldsymptomless seeds whereas in outer and inner (Fig.4) pericarps, endosperm layer (Fig.5) and embryo (Fig. 6) of bolddiscoloured and shrivelled-discoloured seeds in both the samples (Table 1). Conidia of A. alternata were also observed in seed coat of some seeds.

Microtome sections showed mycelium of the pathogen in outer and inner pericarps of all the seed categories (Fig. 7 & 11). In 3 out of 10 bold-discoloured seeds, mycelium penetrated the cuticle and formed small compact knots or cushions in the inner pericarp more abundantly at the hilar end. The cuticle was relatively thin and sinuate with gaps at places. Abundant mycelium was observed in space between endosperm and embryo, between the cotyledons and cells of cotyledons and shoot apex (Fig. 8-10) in severely infected seeds. Thus, A. alternata in sunflower seeds was found to be extra as well as intraembryal in nature. Similar observations were also made by Singh et al.⁸, Raut¹⁵, Kaur et al.¹⁶ and Krishnappa and Shetty.17 Its mycelium in seed coat, endosperm and embryo in infected chilli seeds has also been reported from Rajasthan. In shrivelled- discoloured seeds the parenchyma, aleurone and endosperm layers were completely disintegrated and replaced by mycelium. The embryo was greatly reduced and the cells showed complete depletion of contents.

The presence of mycelium of A. alternata in seeds of sunflower in outer- and

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Fig. 1-6: Infection of Alternaria alternata in sunflower seeds; Fig.1. Seed Categories (left to right) bold-symptomless, bold-discoloured and shrivelled-discoloured seeds. Note greyish brown and brown to black discolouration on seed of later two categories. x 20; Fig. 2. Chain of coindia of fungus on seed surface, x 50; Figs. 3-6. Seed components showing thick, dark, septate, inter- and intra cellular mycelium in cleared wholemount preparations. (3) outer pericarp (4) inner pericarp (5) endosperm and (6) cotyledon. x 125.



Fig. 7-10: Histopathology of infected seeds; Fig. 7. mycelium in seed coat layers. x 25; Fig.8. inter - and intra cellular mycelium in the cotyledonary tissue. x125; Fig. 9. abundant mycelium in space between endosperm and embryo. x 125; Fig. 10. heavy colonization of shoot apex and space between the cotyledons. x 125.



Fig. 11: Semidiagrammatic representation of section of categorised seeds showing expanse of mycelium of Alternaria alternata in sunflower.

- A. Bold- symptomless seed
- B. Bold- discoloured seed
- C. Shrivelled- discoloured seed

(Cot. - Cotyledon; end. - endosperm; hra - hypocotyledonary root axis; ipc - iner pericarp; my- mycelium; opc - outer pericarp; sa - shoot apex; t - testa)

inner pericarp layers of seed coat in boldsymptomless seeds suggests that the pathogen penetrate seed through epidermis mainly towards hilum. The mycelium either remained confined to the layers of seed coat or spread into other tissues in favourable conditions.

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