KINETICS OF CADMIUM UPTAKE IN *STIGEOCLONIUM TENUE* KUTZ.

M. VANAJA*, N.V.N. CHARYULU and K.V.N. RAO

Central Research Institute for Dryland Agriculture, Santoshnagar, Saidabad, Hyderabad 500059 Andhra Pradesh, India.

The tolerance index concentration (TIC) and kinetics of cadmium uptake in *Stigeoclonium tenue* Kutz were studied. The TIC of Cd was 11.49 mg L⁻¹ (10.26 μ M Cd). The uptake of Cd was concentration and time dependent. At low levels (10 μ M and 50 μ M Cd), the absorbed Cd increased till the end of the experimental period. Whereas at higher concentrations of Cd (100 μ M and 150 μ M Cd), the absorbed amount of Cd at the end of the experiment was double when compared to the first hour absorption. Both the adsorption and absorption rates were higher during initial hours of incubation. At lower concentration (10 μ M and 50 μ M Cd), the amount of absorbed Cd was more than the adsorbed Cd at the end of the incubation period. At higher concentrations (100, 150 and 200 μ M Cd) the amount of adsorbed Cd was more than the absorbed Cd. There seems to be a differential mechanism of adsorption and absorption is involved in the tolerance of *S. tenue* to higher concentrations of Cd.

Keywords: Cadmium; Heavy metals; Stigeoclonium tenue Kutz; Tolerance index concentation.

Introduction

Industrialization, urbanization and anthropogenic activities resulted in the environmental pollution by heavy metals¹. The effect of a variety of metals on physiological and biochemical functions of different plant systems has been reviewed2-7. Algae growing in waters polluted by heavy metals have been shown to develop tolerance to higher concentration of heavy metals than those growing in unpolluted regions⁸⁻¹⁰. Stigeoclonium tenue has been reported as an important inhabitant of organically polluted rivers¹¹⁻¹³. Several authors suggested methods to use Stigeoclonium as a bioassay organism for pollution¹⁴⁻¹⁵. After mercury and copper, cadmium has been reported to be the most toxic element to plants¹⁸. Cadmium has been shown to inhibit growth¹⁹, photosynthesis²⁰, reduce the content of photosynthetic pigments²¹, cause damage to the structure of chloroplast²² and mitochondria²³.

Stigeoclonium tenue Kutz. isolated from the effluents of Ramagundam thermal power plant (Andhra Pradesh) has been used in the present investigation to study the tolerance and the kinetics of cadmium uptake. This alga has been used to understand various aspects of cadmium toxicity²⁴⁻²⁵, tolerance and defense mechanisms developed by tolerant species. This would be useful for developing bio-treatment system for remedial of heavy metal pollution.

Material and Methods

Stigeoclonium tenue Kutz. isolated from the Ramagundam thermal power plant effluents was grown in pure anxenic cultures²⁵ on modified Bold's meium²⁶. These cultures were maintained at $18 \pm 1^{\circ}$ C with cool white florescent light intensity of 6000 lux. Cadmium was given in the form of cadmium suphate (3 CdSO₄. 8H₂O). Pure bacteria free inoculum of the alga was taken from the stock cultures. The inoculum consisted of an apparent homogenous suspension of the broken filaments of the alga to facilitate the transfer of equal amount of inoculum for all treatments.

Toxicity test and determination of tolerance index concentration (TIC): To study the toxic effect of Cd and to determine TIC, *Stigeoclonium* was grown on media with Cd concentration ranging from 1.0 mg 1^{-1} , with an increment of 1.0^{-1} . The basal medium was used as control. For each treatment, ten tubes were maintained under standard culture conditions. The toxic and tolerance index concentration (TIC) for Cd was calculated following the semi-quantitative scale as per Harding and Whitton²⁷.

The growth of the alga at different

Cd concentrations was compared visually under microscope with control on 6th, 8th and 14th day after re-inoculation. At the end of 14th day(incubation period), the concentration of Cd at which the growth of the alga was as normal and vigorous as that of the control was categorized as "maximum concentration causing no inhibition of growth (1)". At the end of 8th day, the concentration of Cd at which slight inhibition of growth occurred, the alga from that concentration was re-inoculated to fresh basal medium and grown up to 14th day. The minimum concentration at which re-inoculated alga grow as fast as control was categorized as "minimum concentration causing the slight inhibition of growth (2)". The concentration of Cd at which the re-inoculated alga showed even slight recovery by 14th day was recorded as the "maximum concentration at which the alga is alive (3)". The concentration of Cd where there was no recovery of the alga and death of the cells at the end of 14th day was considered as the "minimum concentration at which the alga was killed (4)". While determining these concentrations for different categories, the identified concentration range was further narrowed by taking 0.1 to 0.9 mg 1⁻¹ of that particular concentration.

Based on these observations the non-inhibitory, lethal and tolerance index concentration (TIC) were calculated. The "non-inhibitory concentration" is the geometric mean of the maximum concentration causing no inhibition (1) and minimum concentration causing slight inhibition (2). The "lethal concentration" is the geometric mean of the maximum concentration at which the alga was alive (3) and the minimum concentration. at which the alga was killed (4). The tolerance index concertation (TIC) is th geometric mean of non-inhibitory and lethal concentrations of Cd.

Determination of the cellular cadmium in the algal mat was carried out as per Singh and Yadava²⁸ by atomic absorption spectrophotometer (AAS- Prekin-Elmer 2380) at 228.8 nm. To measure the adsorbed cadmium, the algal mat was washed with 0.2 M cysteine solution. The washings were pooled and analysed for Cd by AAS. The amount of cellular (absorbed) Cd and adsorbed cadmium was expressed as μg of Cd per gram dry weight. Each value is a mean of three replicates and all the results were statistically analysed.

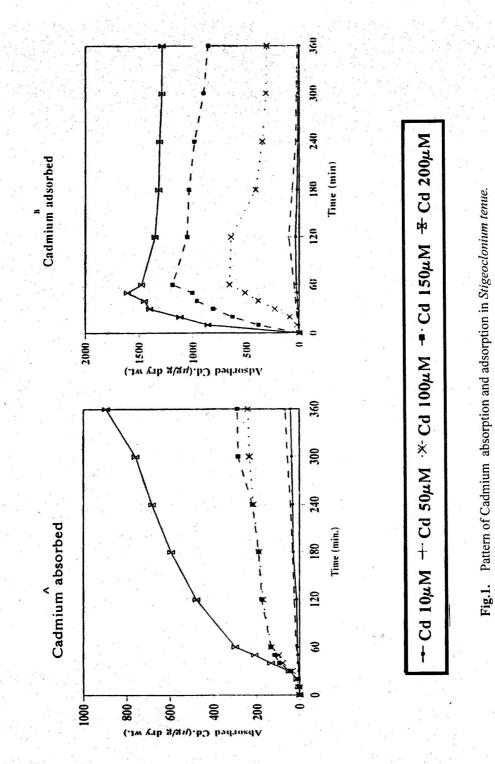
Results and Discussion

Studies on Cd toxicity and TIC have shown that S. tenue showed no inhibition of growth upto 6.2 mg 1⁻¹ (55.18 µM) Cd wherein it grew as fast as the control. At 6.8 mg1-i (60.52 µM) and above Cd inhibited the growth of the alga. The alga taken out form this concentration (i.e.6.8 mg 1-1 or 60.52 µM) on 8th day and re-inoculated in the basal medium showed a lag phase in the beginning, but recovered and showed better growth up to 14th day. The alga grown at 18.2 mg1-1 (161.98µM) and taken out at 8th day recovered slightly after re-inoculation in the basal medium. But the growth was not as good as the control. When the alga was grown at 22.7 mg 1⁻¹ (202.03 μ M) Cd the growth was completely inhibited and finally resulted in the death of the alga. From the toxicity and TIC studies the following observations were also made.

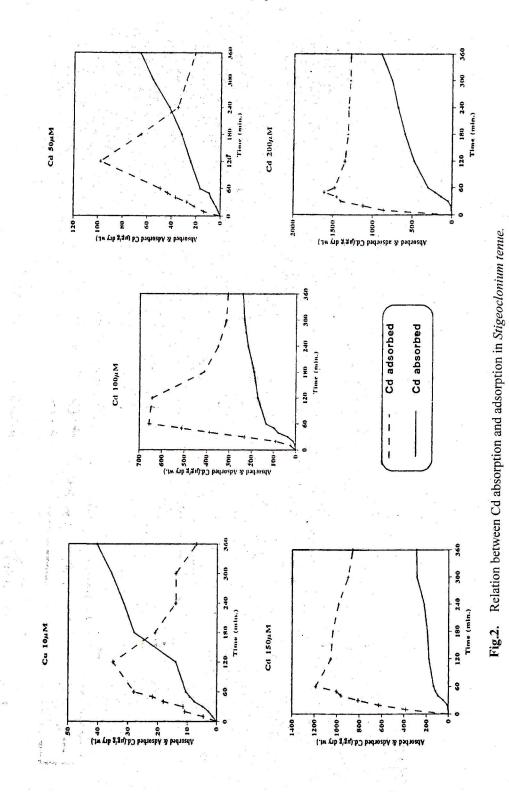
 $i. \mbox{ Non-inhibitory concentration of Cd} & 6.49 mg1^4 \ (57.76 \ \mu M \ Cd) \\ ii. \ Lethal \ concentration \ of Cd & 20.33 \ mg1^4 \ (180.94 \ \mu M \ Cd) \\ iii. \ Tolerance \ index \ concentration \ (TiC) & 11.49 mg1^4 \ (10.26 \ \mu M \ Cd) \\ }$

The present investigation has shown that *S. tenue* can withstand high Cd concentration and can grow normally up to about 55 μ M Cd. This observation is supportive of the classification of this alga as a chemobiont by Fjerdingstad²⁹. However there are no reports regarding the resistance of this alga to Cd.Harding and Whitton²⁷ showed *Stigeoclonium* to be tolerant to high zinc levels. *S. tenue* tolerance to zinc and lead³⁰, copper and chromium ²⁹⁻³¹ have been reported. Based on the present study on tolerance index concentration (TIC) for cadmium, *Stigeoclonium* can be categorized as resistant to high levels of cadmium.

The amount of Cd absorbed or cellular Cd increased with the increase in Cd concentration (Fig.1) and the uptake of Cd are time dependent. The rate of absorption



J. Phytol. Res. 13 (2) : 117-122, 2000



120

in the first one hour was more at all levels of Cd, when compared with later absorption rates (Fig.1). At 10and 50 μ M Cd, the absorbed Cd increased till the end of the experimental period, and it was four fold when compared to the absorption at the end of first one hour. Whereas at 100 and 150 μ M Cd, the amount of absorbed Cd was doubled when compared to the first hour absorption. At the end of the experiment the algal cells absorbed 40.42 μ g g⁻¹ dry weight at 10 μ M Cd, and it increased by about 6 fold at 100 μ M and by 22 fold at 200 μ M Cd (Fig.1).

The amount of adsorbed Cd at initial periods of incubation was more than absorbed Cd at all concentrations of cadmium. In the first two hours of incubation, the adsorbed .Cd at 10 and 50 μ M Cd level was much higher than absorbed Cd. However, at later hours of incubation (as time lapsed), the content of adsorbed cadmium decreased to less than absorbed Cd (Fig.2). With the increase in the Cd concentration in the medium (100,150 and 200 µMCd), maximum amount of Cd adsorption reached within 50 to 60 minutes (Fig.2). The amount of adsorbed Cd at the end of the experimental period remained higher than the absorbed Cd at these higher concentrations of cadmium. The studies on the kinetics of Cd uptake have shown that:

- Both absorption and adsorption rates were high during the initial hours of incubation.
- (ii) At low Cd levels (10and 50 μM Cd), the amount of absorbed Cd was more than the adsorbed Cd at the end of incubation period.
- (iii) At higher Cd levels (100, 150 and 200 μ M Cd) the amount of adsorbed Cd was more than absorbed Cd at the end of incubation period.

The metal accumulation by microorganisms generally comprises of two phases; a rapid binding of cations to the negatively charged groups on the cell surface and the subsequent metabolism dependent intra-cellular uptake³². The present study has shown that Stigeoclonium cells removed cadmium ions from the medium in two possible ways. Firstly, the metal was adsorbed on the surface of the algal cells, mostly through passive uptake as shown in the cysteine washings. Secondly, a large part of Cd was absorbed through energy dependent active uptake. This pattern of Cd absorption and adsorption was dependent on the amount of Cd in the external medium and also on the length and/or duration of exposure of the algal cells to metal ions (Fig.1). At low levels of Cd, the adsorption of Cd was more at initial period than the absorbed Cd. At later stages of incubation, the amount of Cd absorbed has over taken the amount of Cd adsorbed (Fig.2). Similar biphasic pattern of metal uptake has also been reported in other microbes 32-34 and higher plants35.

Acknowledgements

We thank Head, Botany Department, Osmania University for providing facilities. M. Vanaja is grateful to CSIR for the award of fellowship during the tenure of this work.

References

- 1. Foy C D, Chaney R L and White M C 1978, Ann. Rev. Plant Physiol. 29 511
- 2. Haug A 1984, CRC Crit. Rev. Plant Sci. 1 345
- Haug A and Caldwell C R 1985, In: Frontiers in Membrane Research in Agriculture. Totowa, New Jersey, Rowman and Allanheld, pp 359
- 4. Steffens J C and Williams W 1987, *Plant Biol.* 4 109
- 5. Blum A 1988, *Plant Breeding for Stress Environments*. Boco Raton Fla CRC Press.
- Cumming J R and Taylor GJ 1990, In: Stress Responses in Plants: Adaptaion and Acclimation Mechanisms. Wiley-Liss Inc. New York. pp 329
- Prasad M N V 1997, In : *Plant Ecophysiology*, John wiley & Sons. Inc. pp. 207
- 8. Bryan G W 1971, Proc. R. Soc. Lond. B 177 389
- 9. Stokes P M, Huchinson T C and Krauter K 1973, Can. J. Bot. 51 2155
- 10. Agrawal M and Kumar H D 1975, *Ind. J. Ecol.* **2** 94
- 11. Hynes HBN 1960, *The Biology of Polluted Water*. Liverpool University Pres, Liverpool.
- 12. McLean RO and Benson-Evans 1974, Br. Phycol. J. 9 83
- 13. Venkateswarlu V 1981, In: WHO workshop on biological indicators and indices of environmental pollution.

122

Vanaja et al.

- Trotter DM, Hendricks AC and Cairns J Jr. 1978, Wat. Res. 12 185
- 15. De Vries P J R, Torenbeek M and Hillebrand H 1983, Aquat. Bot. 17 95
- 16. De Vries P J R and Kamphof G J 1984, Br. Phycol. J. 19 349
- De Vries P J R and Hotting E J 1985, *Wat Res.* 19 1405
- 18. Sorentino C 1979, Phykos 18 149
- Hutchinson T C and Stokes P 1975, In: Water Quality Parameter. Philadelphia, American Society for Testing of materials, Special Technical Publication No. 573: 320
- 20. Stratton G W and Corke C T 1979, *Chemosphere* 5 277
- 21. Conway H L 1978, J Fish Res. Bd Canada 35 286
- 22. Visviki I and Ranchlin J W 1992, Arch. Environ. Contam. Toxicol. 23 420
- 23. Silverberg BA 1976, Phycologia 15 155
- Vanaja M 1995, Studies on the mechanisms of cadmium tolerance in Stigeoclonium tenue Kutz. Thesis, Osmania University, Hyderabad.
- 25. Vanaja M, Charyulu NVN and Rao KVN 1989,

Proceedings of National Science Academy B55 (5-6) 489

- Bischoff H W and Bold H C 1963, In : University of Texas Publ. 6318 Austin, Texas
- 27. Harding JPC and Whitton B A 1976, *Br. Phycol. J.* 13 417
- Singh S P and Yadava V 1985, J. Gen. Appl. Microbiol. 31 39
- 29. Fjerdingstand E 1965, Int. Rev, ges. Hydrobiol. 50 475
- 30. McLean R O 1974, Br. Phycol. J. 9 91
- 31. Pallmer C M 1959, Publication of US Public Health Service 657: 1-88.
- 32. Norris P R and Kelly DP 1977, *J. Gen. Microbiol.* 99 317
- Sakaguchi T, Tsuji T, Nakajima A and Horikoshi T 1979, European J Appl. Microbial Biotechnol. 8 207
- 34. Khummongkol D, Canterford GS and Fryer C 1982, *Biotech. Bioengg.* XXIV 2643
- Macek T, Kotrba P, Suchova M, Skacel F, Demanerova K and Rumi T 1994, *Biotch. Lett.* 16 621