

EFFECT OF PARTICULATE POLLUTION ON *PISUM SATIVUM* L.

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The effect of particulate pollution on morphology and biochemistry of *Pisum sativum* was studied. The results were compared with non-polluted plants. In polluted plants, there is a reduction of 51.89% in height of shoot, 62.05% in phytomass and 64.10% in yield. Chlorophyll content in polluted plants exhibit a reduction of 38.32%. Stomatal index and trichome frequency increases and stomatal frequency decreases in polluted plants.

Keywords : Particulate pollution; Phytomass; Chlorophyll; *Pisum*.

Introduction

The deteriorating quality of the environment is causing worldwide concern and mankind is facing newer and unimaginable kinds of environmental problems. The present work is an attempt to record the response of *Pisum sativum* to cement kiln dust (particulate) pollution.

Materials and Methods

Pisum sativum L. seeds were obtained from Gujarat Agricultural University. Seedlings were raised in University Botanical Garden in two separate plots. The plants in one plot served as control (non-polluted), whereas those in the other were uniformly dusted with cement kiln dust (pollu-

ted). Growth analysis and biochemical estimations were carried out in non-polluted and polluted plants, simultaneously. The Methods followed for various estimations were : Chlorophyll content—Mac Lachlan and Zalik (1963); Protein—Layne (1957); Starch—Mc Credy *et al.* (1950); Lipid—Folch *et al.* (1957) and Bragdon (1951); Amino acid—Moore and Stein (1948); Total sugars—Dubois *et al.* (1956); Reducing sugars—Miller (1972); Total phenols—Bray and Thorpe (1954); O.D. phenol—Arnow (1937). Epidermal peels were taken from the leaflets by direct peel method and stained with Delafield's haematoxylin. Stomatal index was

calculated as defined by Salisbury (1927, 1932).

Results and Discussion

In polluted plants, there is a reduction of 51.89% in height of shoot; 33.33% in root length; 50% in number of leaflets and 62.05% in phytomass. There is an increase of 41.18% in root/shoot ratio of polluted plants. There is 66.67% and 61.10% of reduction in numbers of flowers and fruits, respectively, in polluted plants (Table 1).

Chlorophyll content exhibited a reduction of 32.16% in polluted plants. In polluted plants, there is a reduction of 45.59% protein, 21.94% starch, 24.05% total sugar, 24.79% reducing sugars, 41.30% lipid, 28.33% amino acids, 37.62% total phenols and 33.51% O.D. phenol contents (Table 2).

Epidermal cells are polygonal with wavy anticlinal walls. A reduction is observed in stomatal frequencies, while stomatal index and trichome frequency increased in polluted plants. Leaves are amphistomatic with anomocytic stomata as the dominant type on both surfaces in non-polluted and polluted plants. Stoma with single subsidiary cell is common in non-polluted and polluted plants. However, in polluted plants its percentage increases along with abnormal stomatal types like stoma

with single guard cell and contiguous stomata etc. Only eglandular trichomes were observed only on abaxial surface of both non-polluted and polluted plants (Table 3).

Schonbeck (1960) observed that physiological balance in affected plants were altered by dust increasing susceptibility to infection. In the present study, cement kiln dust was found to reduce shoot length. The inhibition in growth might be attributed to the reduced intensity of light energy available for photosynthesis through coating of leaves (Indhirabai *et al.*, 1988, 1989). The root length was reduced in polluted plants along with the reduction in nodulation up to 60%. This might be due to increased pH of soil. The number of branches in polluted plants was found to be decreased due to decrease in shoot length. Appreciable reduction in shoot length and number of branches in polluted plants results in considerable decrease in number of leaflets per plant (Darely, 1966).

The phytomass and net primary productivity values of cement kiln dust polluted plants show decreased values. Darely (1966) and Lerman (1972) noted that the reduction in phytomass is due to interruption in gaseous exchanges on a result of stomatal clogging which leads to reduction in photosynthesis. The root/shoot ratio values in polluted

Table 1. Morphological parameters of non-polluted and polluted plants (average of five replicates)

| age of plant (days) | Height of shoot (cm) | Length of root (cm) | Number of leaves | Total phyto-mass (gm) | NPP gm/plant/day | Root shoot ratio | Number of flowers | Number of fruits |
|---------------------|----------------------|---------------------|------------------|-----------------------|------------------|------------------|-------------------|------------------|
| 20 | NP 40 | 9 | 9 | 1.860 | 0.093 | 0.23 | 3 | — |
| | P 35 | 8 | 7 | 1.322 | 0.066 | 0.23 | 2 | — |
| 40 | NP 55 | 13 | 21 | 9.778 | 0.244 | 0.24 | 17 | 4 |
| | P 42 | 10 | 18 | 5.529 | 0.138 | 0.24 | 10 | 2 |
| 60 | NP 112 | 15 | 170 | 19.613 | 0.327 | 0.13 | 30 | 19 |
| | P 65 | 10 | 68 | 9.578 | 0.160 | 0.15 | 19 | 8 |
| 80 | NP 162 | 18 | 250 | 53.530 | 0.670 | 0.11 | 85 | 40 |
| | P 80 | 14 | 120 | 20.358 | 0.254 | 0.18 | 40 | 15 |
| 100 | NP 185 | 19 | 180 | 40.157 | 0.402 | 0.10 | 12 | 78 |
| | P 89 | 15 | 90 | 15.240 | 0.152 | 0.17 | 4 | 28 |

Table 2 : Biochemical parameters of non-polluted and polluted plants (average of five replicates)

| Age of plant (days) | | Total chloro-phyll content | Protein content mg/gm | Starch content mg/gm | Total sugar mg/gm | Reducing sugar mg/gm | Lipid content mg/gm | Amino acid content mg/gm | Total phenol content mg/gm | O.D. phenol content mg/gm |
|---------------------|----|----------------------------|-----------------------|----------------------|-------------------|----------------------|---------------------|--------------------------|----------------------------|---------------------------|
| 20 | NP | 0.977 | 0.115 | 0.961 | 5.431 | 1.142 | 0.646 | 0.715 | 0.786 | 0.293 |
| | P | 0.782 | 0.098 | 0.890 | 4.686 | 0.896 | 0.518 | 0.621 | 0.642 | 0.221 |
| 40 | NP | 1.949 | 0.152 | 1.544 | 11.140 | 3.514 | 1.032 | 1.038 | 0.981 | 0.410 |
| | P | 1.434 | 0.115 | 1.350 | 9.117 | 2.257 | 0.729 | 0.786 | 0.765 | 0.295 |
| 60 | NP | 2.828 | 0.188 | 2.575 | 16.493 | 7.148 | 1.536 | 2.348 | 1.238 | 0.550 |
| | P | 1.896 | 0.126 | 1.909 | 12.280 | 3.999 | 1.022 | 1.526 | 0.927 | 0.377 |
| 80 | NP | 3.265 | 0.242 | 3.728 | 18.232 | 8.412 | 2.121 | 4.251 | 1.627 | 0.743 |
| | P | 2.014 | 0.151 | 2.595 | 13.307 | 5.754 | 1.324 | 2.178 | 1.015 | 0.494 |
| 100 | NP | 1.281 | 0.193 | 1.121 | 15.341 | 7.368 | 2.421 | 7.171 | 1.815 | 0.908 |
| | P | 0.869 | 0.105 | 0.875 | 11.652 | 5.541 | 1.421 | 5.064 | 1.521 | 0.661 |

NPP = Net primary productivity; NP = Non-polluted; P = Polluted.

Table 3 : Empidermal features of non-polluted and polluted plants (average of five replicates)

| | STOMATA | | | | TRICHOME | | | | | | | |
|----|--------------------------|----------------|----------------|------------|--------------------|----------------------|-----|----------|----------------------|----------|-----|---|
| | Epidermal cell frequency | Stomatal index | Stomatal types | | Trichome frequency | Eglandular trichomes | | | | | | |
| | | | Para-cyctic | Anomocytic | | | | Diacytic | Stomatal single S.C. | Abnormal | | |
| Ad | 726 | 18.6 | 166 | 6.0 | — | 73.0 | 3.0 | 18.0 | — | — | — | — |
| NP | 396 | 23.4 | 121 | 5.0 | — | 66.0 | 2.0 | 22.2 | 4.8 | — | — | — |
| Ab | 730 | 15.8 | 137 | 5.6 | 4.0 | 67.0 | 4.3 | 19.1 | — | 28 | 100 | — |
| NP | 488 | 18.4 | 110 | 7.3 | 3.0 | 61.4 | 2.3 | 24.8 | 1.2 | 33 | 100 | — |

Ad = Adaxial; Ab = Abaxial; NP = Non-polluted; P = Polluted.

plants were always higher than non-polluted plants (Singh and Rao, 1981). Number of flowers and number of fruits per plant exhibited a reduction in polluted plants. The cement kiln dust polluted microenvironment provides unfavourable ecological conditions for pollen germination and fertilization (Czaja, 1962). Borka (1980) stated that the damaging effects of cement kiln dust were not only a reduction in vegetative parts, but also a reduction in the formation of reproductive organs and consequently in fertilization.

One of the most characteristic biochemical features of cement kiln dust polluted plants is a reduction in the total chlorophyll content, as reported by a number of workers (Czaja, 1962; Dorely, 1966; Borka, 1986). The reduction in chlorophyll content is due to the influence of pollution (Gilbert, 1978). The absorption of soluble portion of cement kiln dust into leaf tissue and damage to the chloroplasts leads to a reduction in total chlorophyll content (Singh, 1980).

Present observation on the reduction in protein content in polluted plants is parallel to the results of many workers (Pawer *et al.*, 1982; Mandre and Kangur, 1986). It thus appears that the total protein content is also a suitable indicator of particulate pollution level.

Ting and Mukerji (1971) reported a reduction in total sugar content in polluted plants which may be due to reduced photosynthetic rate. Lipid content decreases in polluted plants (Grunwald, 1981). The direct relation between the phenolic content and age of plant indicates that this parameter could serve as an index of increasing pollution load.

Increase in stomatal frequency in polluted plants is reported by Chakraborty and Gupta (1981) and Jafri *et al.* (1979). Sharma and Butler (1973) suggested that the high density of trichomes in polluted plants may be a protection mechanism to prevent direct exposure of leaf parts to sun rays, thus lowering the leaf temperature and reducing the rate of some metabolic reactions.

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References

- Arnou L E 1937, *J. Biol. Chem.* **118** 531
- Borka G 1980, *Environ. Pollut.* **22** 75
- Borka G 1986, *Acta. Agr. Hung.* **30** 289
- Bragdon J H 1951, *J. Biol. Chem.* **190** 513
- Bray H G and Thorpe W V 1954, *Meth. Biochem. Anal.* **1** 27

- Chakraborty T and Gupta D 198 , *Indian Acad. Sci. (Pl. Sci.)* 90 305
- Gzaja A T 1962, *Staub.* 22 228
- Darely E F 1966, *J. Air Pollu', Contr. Ass.* 16 145
- Dubois M, Gilles K A, Hamilton J K, Roberts P A and Smith F 1956, *Analyt. Chem.* 28 350
- Folch J, Less M and Stanley G H S 1957, *J. Biol. Chem.* 226 497
- Gilbert O L 1978, *New Phytol.* 67 15
- Grunwald C 1981, *Pl. Pysiol.* 68 867
- Indhirabai K, Dhanalakshmi S and Lakshmanan K K 1988, *APSI* 1 91
- Indhirabai K, Dhanalakshmi S and Lakshmanan K K 1989, *Geobos* 16 189
- Jafri S, Srivastava K and Ahmad K J 1979, *Ind. J. Air Pollut Control* 2 74
- Layne E 1957 *Methods in Enzymology* Academic press, New York
- Lerman S L 1972, Ph. D. Dissertation, University of California
- Mac Lachlan S and Zalik S. 1963, *Can. J. Bot.* 41 1053
- Mandre M and Kangur A 1986, *Proc. of the Acad. of Sci. of the Estonian SSR* 35 1222
- Mc Creddy R M, Guggloz J, Silvieira V and Owens H S 1950, *Anal. Chem.* 22 1156
- Miller G L 1972, *Anal. Chem.* 31 426
- Moore S and Stein W H 1948, *J. Biol. Chem.* 176 367
- Pawar K, Trivedi L and Dubey P S 1982, *Int. J. Environ, Studies* 19 221
- Salisbury E J 1927, *Phil. Trans. R. Soc. B.* 26 1
- Salisbury E J 1932, *Beih. Bot. Zentralb.* 49 408
- Schonoback H 1960, *Nordisk. Westfol. Bochum* 89
- Sharma G K and Butler. J. 1973, *Environ. Pollut.* 5 287
- Singh S N 1980, *J. Exp. Bot.* 31 1701
- Singh S N and Rao D N 1981, *Ind. J. Environ. Hlth.* 20 258
- Ting J P and Mukerji S K 1971, *Am. J. Bot.* 58 497.